

Hastane İnfeksiyonları ve Mikrobiyoloji Laboratuvarı

Barış Otlu
İnönü Üniversitesi Tıp Fakültesi
Tıbbi Mikrobiyoloji Anabilim Dalı



Hastane İnfeksiyonları Tarihi

3. Yüzyıl Roma Şiiri, Bir hastanın Dr. Simmakus'a seslenişi

“Hasta oluyordum ve sen hemen geldin
Yanında yüz öğrenciyle, ah **Simmakus**
Yüz soğuk el bana dokundu
Hiç ateşim yoktu, ah Simmakus, şimdi var”



Hastane İnfeksiyonları Tarihi

- Hastane, hastane infeksiyonları ve mikrobiyolojik tanı kavramlarının birlikte geliştiği düşünülebilir.



18.yy' da Hotel Dieu;

ampütasyonlardan ölüm oranı %60

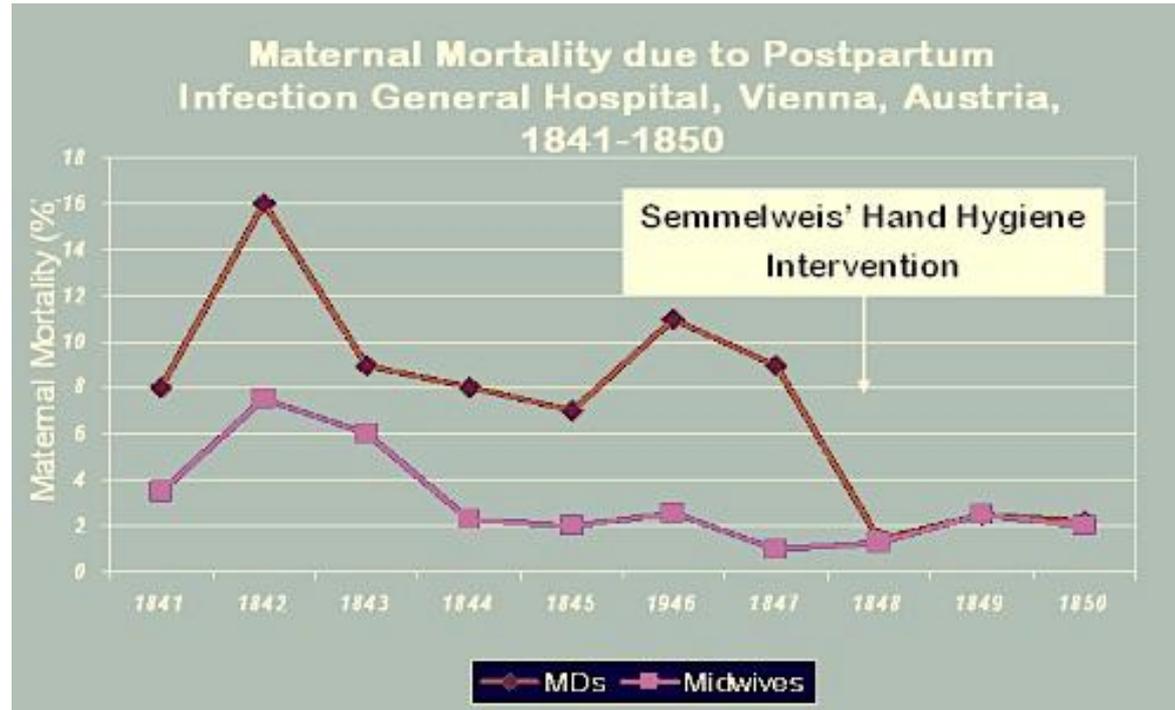
salgın sırasında lohusalık hummasından ölüm oranı 19/20

Hastane İnfeksiyonları Tarihi

- **1840** Hastane İnfeksiyonlarının tespiti ve önlenmesi ile ilgili ilk bilimsel yaklaşım.

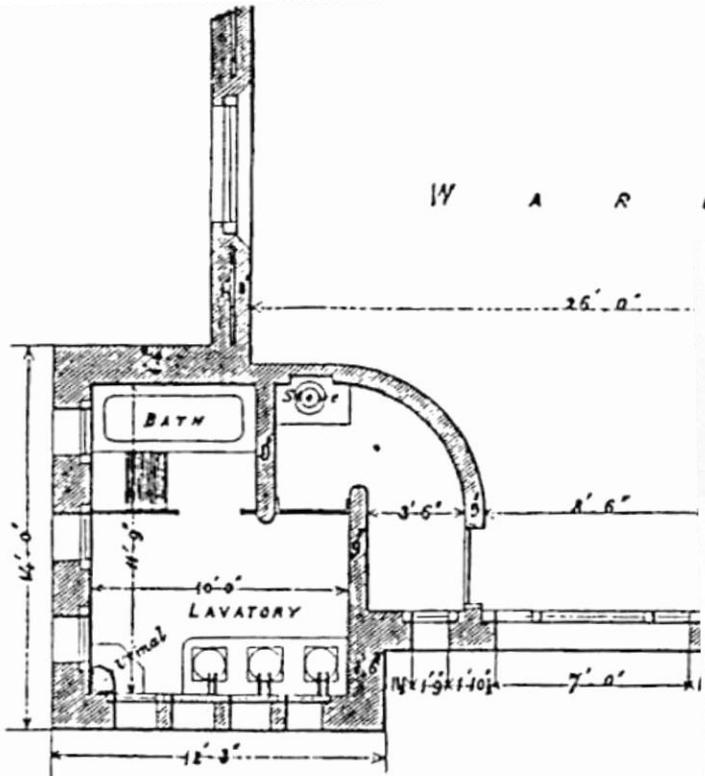


Ignaz Semmelweis



Hastane İnfeksiyonları Tarihi

from *Notes on Hospitals* published in 1863



Plan of Lavatory, Baths, and Water-closet

from

HOSPITAL GENERAL STATISTICAL FORM.

This Sheet will serve for the Classification of Cases in Hospitals under the following headings:—"Remaining, 1st January"—"Admitted"—"Cured (or Relieved)"—"Dead"—"Discharged incurable, for Irregularities, or at their own Request"—"Remaining, 31st December"—"Duration of Cases in Days."

Write the Name of Hospital, the Sex, the required Heading, and Date, with the Pen.

Age	MONTHS												98 and upwards	Total											
	0	1	2	3	4	5	10	15	20	25	30	35			40	45	50	55	60	65	70	75	80	85	90
CLASS I.—ORDER I. (ZYMOTIC DISEASES.)																									
Small Pox																									
Measles																									
Whooping Cough																									
Croup																									
Scarlatina																									
Diphtheria																									
Typhus, Catarrh, Influenza																									
Typhoid fever (parvula)																									
Erysipelas																									
Malaria (intermittent)																									
Typhus																									
Hospital gangrene																									
Carbuncle, Ulcer																									
Dysentery																									
Diarrhoea																									
Cholera																									
Typhoid Fever (typhus)																									
Typhus																									
Relapsing Fever (typhus)																									
Ague																									
Remittent Fever																									
Remittent																									
OTHERS																									
ORDER II.																									
Gonorrhoea																									
Primary Syphilis																									
Secondary Syphilis																									
Tertiary Syphilis																									
OTHERS																									
ORDER III.																									
Scarry																									
Furuncul																									
Alcohol																									
a. Delirium tremens																									
b. Intemperance																									
OTHERS																									
ORDER IV.																									
Thrush																									
Furrow																									
Warts																									
Worms																									
OTHERS																									
CLASS II.—ORDER I. (CONSTITUTIONAL DISEASES.)																									
Joint																									
Dropsy																									
Labour																									
Cancer (benign)																									
Mortification																									
OTHERS																									
ORDER II.																									
Scrofula																									
Tuberculosis																									

Most of the diseases in the following marginal list are of less frequent occurrence in hospitals; and all, with some exceptions, are classed as "Others" in their respective orders in the left hand column. They will be distinguished in abstracting the disease by writing the age of the person attacked, cured, or dead, &c., against the particular disease in the margin below. Thus, a person aged 40 would, if admitted for "measles," be indicated in the body of this sheet by a dot against "Others" of Class I. Order I.; and his age 40 would be written against "measles" in the margin. And so of other diseases. The diseases not found printed in the margin must be written in their proper compartments. A summary of the facts in the margin should be given in an appendix to the general Table.

CLASS I.
ORDER I.
varicella
scarlatina
measles
erysipelas
dysentery with abscess of liver gland in dysentery
Typhoid fever
ORDER II.
leprosy (Ordn. elephantiasis)
ORDER III.
glanders
epithelioma
melancholic pustule
decays
Infection by gonorrhoea in dissection or by handling the parts of dead animals
ORDER III.
rickets
bronchitis
erysipelas
erythema
ORDER IV.

Hastane İnfeksiyonları Tarihi-Yoğun Bakımlar

- 1952

Intensive care medicine is 60 years old: the history and future of the intensive care unit

Authors: Fiona E Kelly,^A Kevin Fong,^B Nicholas Hirsch^C and Jerry P Nolan^D

Intensive care is celebrating its 60th anniversary this year. The concept arose from the devastating Copenhagen polio epidemic of 1952, which resulted in hundreds of victims experiencing respiratory and bulbar failure. Over 300 patients required artificial ventilation for several weeks. This was provided by 1,000 medical and dental students who were employed to hand ventilate the lungs of these patients via tracheostomies. By 1953, Bjorn Ibsen, the anaesthetist who had suggested that positive pressure ventilation should be the treatment of choice during the epidemic, had set up the first intensive care unit (ICU) in Europe, gathering together physicians and physiologists to manage sick patients – many would consider him to be the 'father' of intensive care. Here, we discuss the events surrounding the 1952 polio epidemic, the subsequent development of ICUs throughout the UK, the changes that have occurred in intensive care over the past 10 years and what the future holds for the speciality.



Fig 3. An 8-year-old girl being hand ventilated via a tracheostomy.



Fig 1. Coventry alligator iron lung.

Hastane İnfeksiyonları Tarihi-Yoğun Bakımlar

Control of cross-infection in an intensive care unit

By D. M. HARRIS, J. M. ORWIN, J. COLQUHOUN AND H. G. SCHROEDER

*From the Department of Bacteriology and the Intensive Therapy Unit,
Royal Hospital, Sheffield*

conditions which rendered them liable to infection. In groups II and III the proportion of patients admitted on account of primary lung disease or major trauma was much lower than in group I.

Table 2 is an attempt to assess the influence of I.P.P.V. on the acquisition of infection. For the three categories of patient considered (all of whom had been subjected to tracheostomy or endotracheal intubation), the incidence of infection was approximately the same whether I.P.P.V. had been used or not.

Considerable attention is

Causative organism	No. of infections	Site of infection			
		Sputum	Tracheostomy	Wound swab	Urine
Coliforms	39	18	6	5	10
<i>Ps. aeruginosa</i>	21	11	7	2	1
<i>Proteus</i> spp.	9	5	1	1	2
<i>H. influenzae</i>	6	5	1	0	0
<i>Strep. pneumoniae</i>	3	3	0	0	0
<i>Staph. aureus</i>	21	10	4	7	0
<i>Strep. faecalis</i>	4	0	1	3	0
<i>Strep. pyogenes</i>	3	0	0	3	0
<i>Candida</i> spp.	14	3	11	0	0
<i>Clostridium welchii</i>	1	0	0	1	0
Totals	121	55	31	22	13



Plan of the Intensive Therapy Unit at the Royal Hospital, Sheffield.

Yoğun Bakım ve Salgınlar

Journal of Hospital Infection (2009) 73, 355–363



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REVIEW

Acinetobacter: an old friend, but a new enemy

K.J. Towner*

Department of Clinical Microbiology, Nottingham University Hospitals NHS Trust, Queen's Medical Centre, Nottingham NG7 2UH, UK

Available online 22 August 2009

KEYWORDS

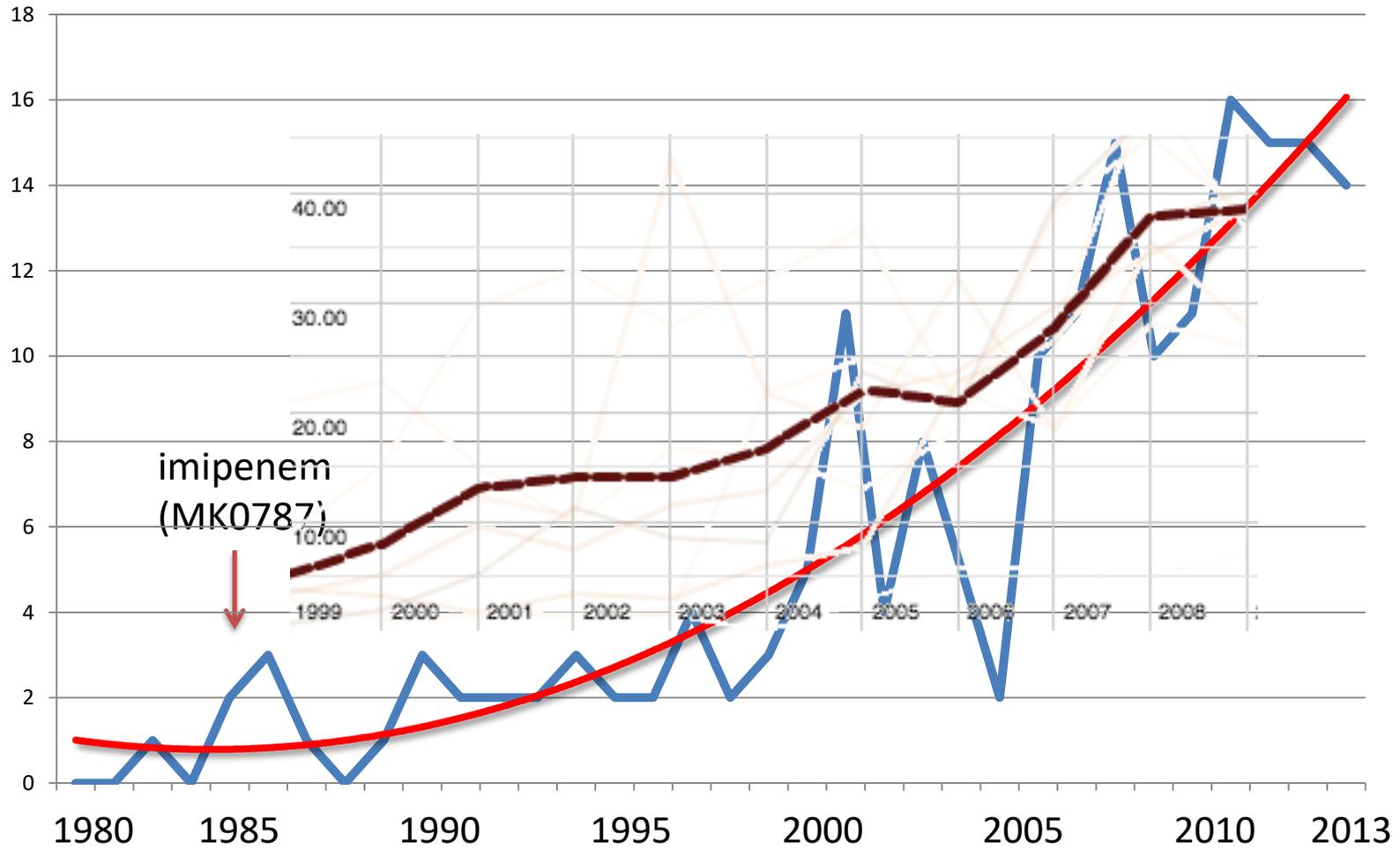
Acinetobacter;
Epidemiology;
Infection control
measures;
Nosocomial infection;
Therapeutic options

outbreaks can occur in such units, involving the infection or colonisation of numerous patients by specific epidemic strains of *A. baumannii*. Recently, a particular problem has concerned cross-infection of injured military patients repatriated from combat regions of the world (e.g. Iraq and Afghanistan). Carbapenems have previously been the treatment of choice for infected patients, but increasing reports worldwide now describe *A. baumannii* isolates resistant to all conventional antimicrobial regimens. Data to support therapeutic use of the limited number of new antimicrobial agents (e.g. tigecycline) with in-vitro activity against these pathogens are still very limited. Detailed advice concerning prevention and control of outbreaks caused by multidrug-resistant strains of acinetobacter is available from the UK Health Protection Agency. In addition to antibiotic prescribing policies and audit, these measures focus on reinforcing standard infection control procedures and precautions, with particular attention to thorough cleaning of patient areas to take account of the long-term survival of acinetobacter after drying and inadequate disinfection. Despite these measures, the problem continues to escalate, with many hospitals worldwide now reporting outbreaks caused by multidrug-resistant strains of acinetobacter.

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PubMed'de son 35 yılda 200 den fazla *Acinetobacter* spp. salgını yayın haline gelmiş !!!



Hastane İnfeksiyonları ve Mikrobiyoloji Laboratuvarı

- İnfeksiyona neden olan etkenin belirlenmesi
 - Etkenin hızla tespiti
 - Antimikrobiyal duyarlılığın belirlenmesi
 - Antibiyotik direnç genlerinin tespit edilmesi
 - Virülans genlerinin belirlenmesi
- İzolatlar arasındaki klonal ilişkinin ve bulaş kaynağının araştırılması
 - Hastane salgınlarının araştırılması
 - Salgının kaynağının belirlenmesi
 - Dirençli bakterilerin izlenmesi
 - Re-aktivasyonun yeni bulaştan ayırt edilmesi

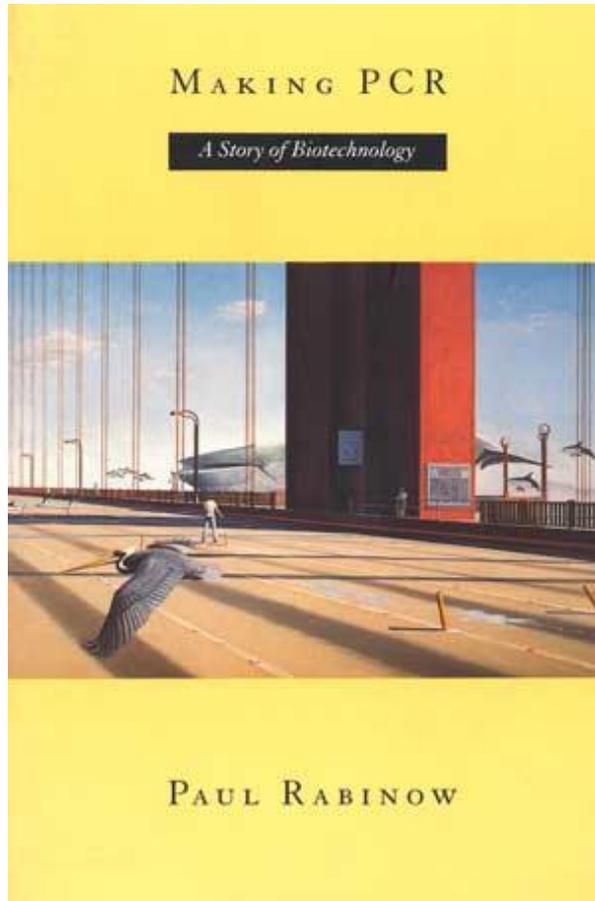
Enfeksiyon etkeninin tespiti

- Etkenin hızla tespiti



Enfeksiyon etkeninin tespiti

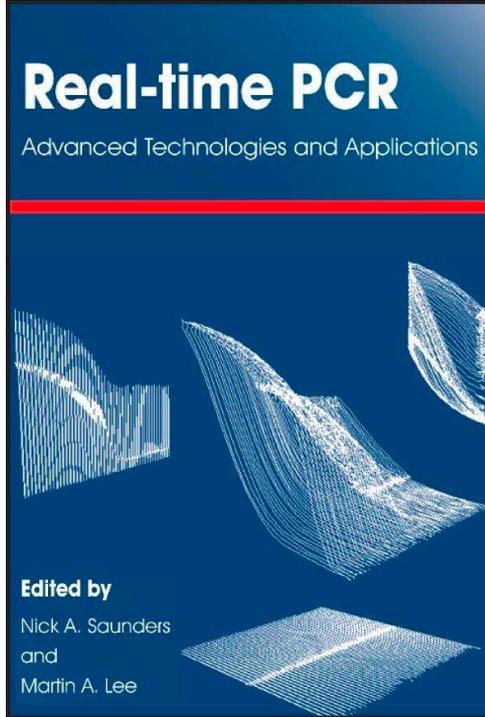
- Moleküler yöntemlerle etkeninin tespiti



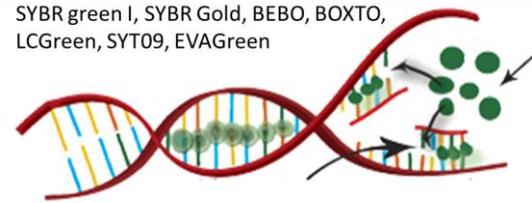
- *PCR*
- *Multiplex PCR*
- *Nested PCR*
- *Seminetested PCR*
- *Broad Range PCR*
- *Hot Start PCR*
- *Touchdown PCR*
- *Reverse Transcription PCR*
- *Real-time PCR*

Enfeksiyon etkeninin tespiti

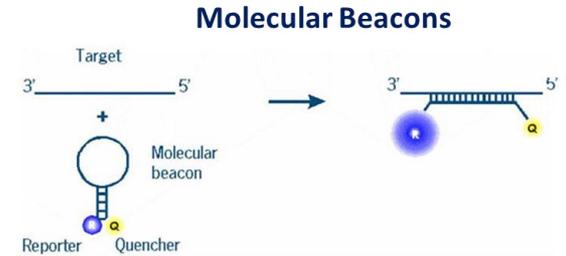
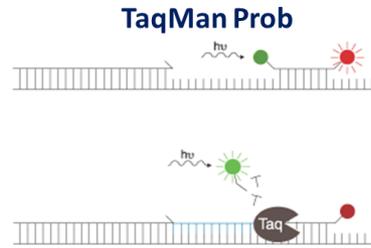
- Moleküler yöntemlerle etkenin tespiti



- Özgül olmayan saptama; florojenik boyalarla



- Özgül saptama; sekans spesifik florojenik oligonükleotid problemlerle



FRET
(Floresans rezonans enerji transferi)



Scorpions Prob



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti
 - Real-time PCR cihazları laboratuvarlarda yerini aldılar.



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti



Healthcare Associated Infections

Xpert MRSA
Xpert SA Nasal Complete
Xpert MRSA/SA SSTI
Xpert MRSA/SA BC
Xpert C. difficile
Xpert C. difficile/Epi
Xpert vanA
Xpert Norovirus
Xpert Carba-R

Critical Infectious Diseases

Xpert MTB/RIF
Xpert Flu
Xpert Flu/RSV XC
Xpert EV

Sexual Health

Xpert CT/NG
Xpert GBS
Xpert GBS LB

Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti



Healthcare Associated Infections

Xpert MRSA

Xpert SA Nasal Complete

Xpert MRSA/SA SSTI

Xpert MRSA/SA BC

Xpert C. difficile

Xpert C. difficile/Epi

Xpert vanA

Xpert Norovirus

Xpert Carba-R

Critical Infectious Diseases

Xpert MTB/RIF

Xpert Flu

Xpert Flu/RSV XC

Xpert EV

Sexual Health

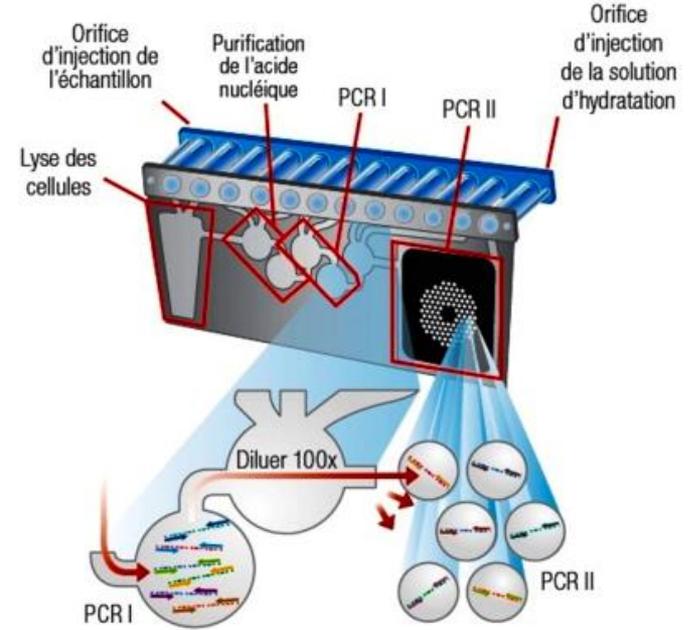
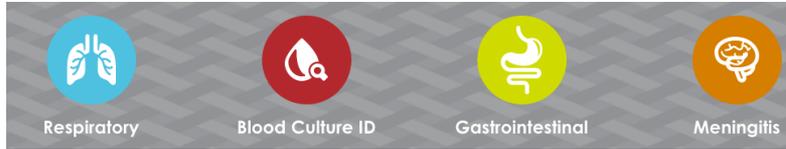
Xpert CT/NG

Xpert GBS

Xpert GBS LB

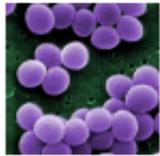
Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti
 - Yüksek multipleks kapasiteye sahip cihazlar (Biofire, Filmarray)



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti
 - Yüksek multipleks kapasiteye sahip cihazlar. Biofire, Filmarray



Gram-Positive Bacteria

Enterococcus

Listeria monocytogenes

Staphylococcus

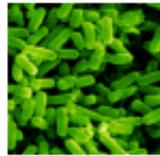
Staphylococcus aureus

Streptococcus

Streptococcus agalactiae

Streptococcus pneumoniae

Streptococcus pyogenes



Gram-Negative Bacteria

Acinetobacter baumannii

Haemophilus influenzae

Neisseria meningitidis

Pseudomonas aeruginosa

Enterobacteriaceae

Enterobacter cloacae complex

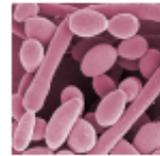
Escherichia coli

Klebsiella oxytoca

Klebsiella pneumoniae

Proteus

Serratia marcescens



Yeast

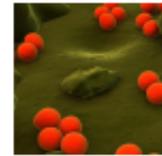
Candida albicans

Candida glabrata

Candida krusei

Candida parapsilosis

Candida tropicalis

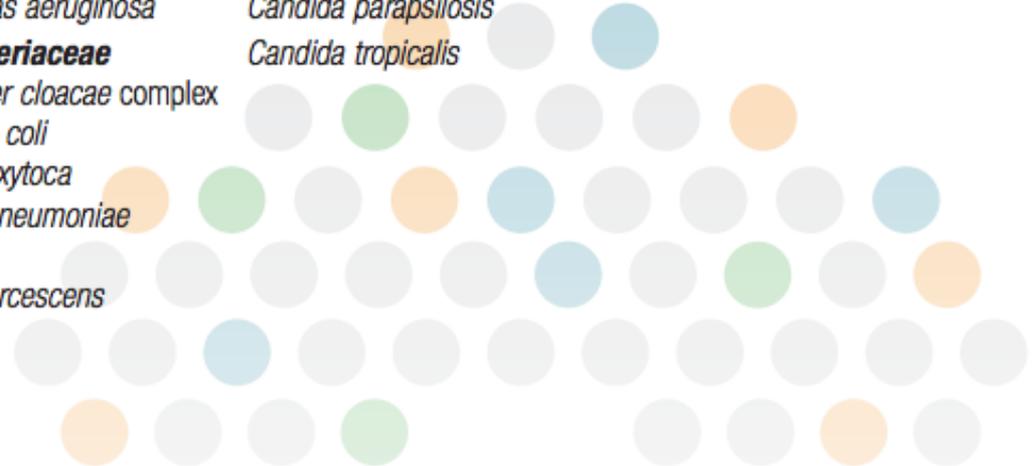


Antibiotic Resistance Genes

mecA – methicillin resistant

vanA/B – vancomycin resistant

KPC – carbapenem resistant



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti

Rapid Detection of Bloodstream Pathogens in Liver Transplantation Patients With FilmArray Multiplex Polymerase Chain Reaction Assays: Comparison With Conventional Methods

B. Otlu^{a,*}, Y. Bayindir^b, F. Ozdemir^c, V. Ince^c, S. Cuglan^a, M. Hopoglu^b, Y. Yakupogullari^a, C. Kizilkaya^d, C. Kuzucu^a, B. Isik^a, and S. Yilmaz^c



Gram negative bacteria	Gram positive bacteria	Yeast	Antimicrobial resistance genes
<i>Acinetobacter baumannii</i>	<i>Enterococcus spp.</i>	<i>Candida albicans</i>	<i>mecA</i> - methicillin resistance gene
<i>Haemophilus influenzae</i>	<i>Listeria monocytogenes</i>	<i>Candida glabrata</i>	
<i>Neisseria meningitidis</i>	<i>Staphylococcus spp.</i>	<i>Candida krusei</i>	<i>vanA/B</i> - vancomycin resistance gene
<i>Pseudomonas aeruginosa</i>	<i>Staphylococcus aureus</i>	<i>Candida parapsilosis</i>	
<i>Enterobacter cloacae</i> complex	<i>Streptococcus spp.</i>	<i>Candida tropicalis</i>	KPC - carbapenem resistance gene
<i>Escherichia coli</i>	<i>Streptococcus agalactiae</i>		
<i>Klebsiella oxytoca</i>	<i>Streptococcus pneumoniae</i>		
<i>Klebsiella pneumoniae</i>	<i>Streptococcus pyogenes</i>		
<i>Proteus spp.</i>			
<i>Serratia marcescens</i>			

Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkeninin tespiti
 - Ortalama kan kültür pozitiflik süresi: 14.5 saat

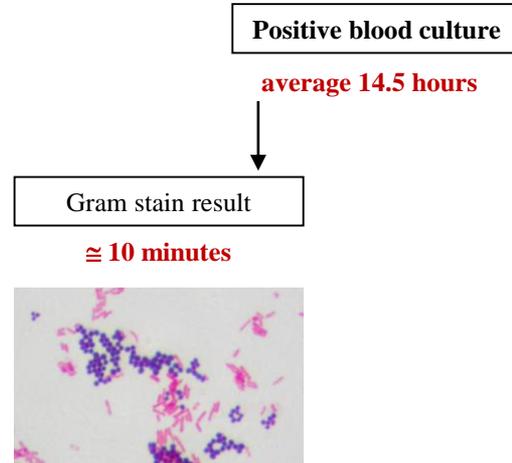


Positive blood culture

average 14.5 hours

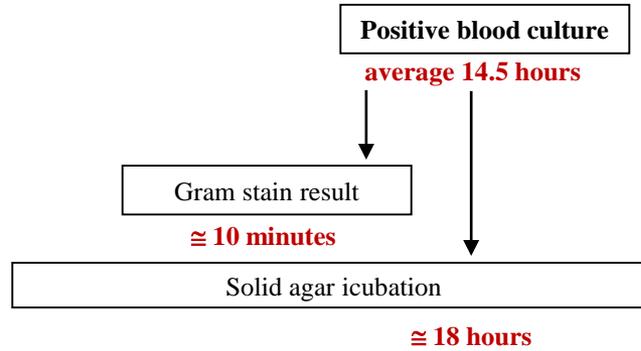
Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Gram boyama



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Kültür sonucu



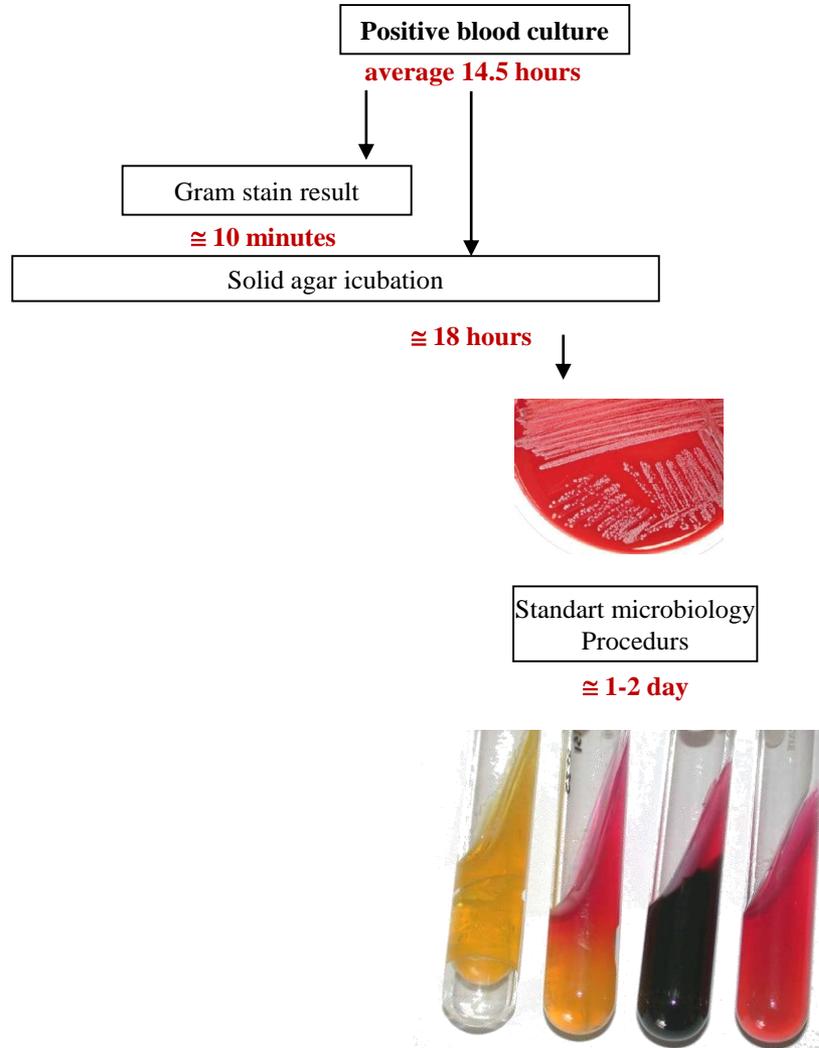
Saf kültür



Subkültür?

Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Kültür sonucu ve tanımlama



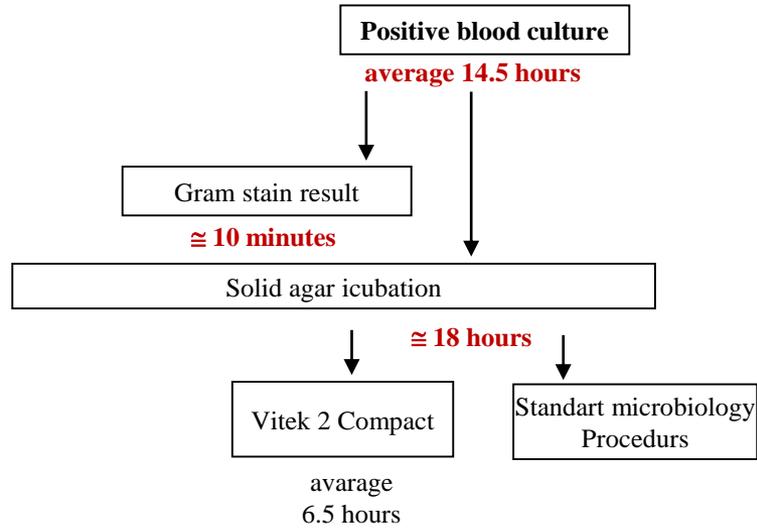
Enfeksiyon etkenin tespiti

- Moleküler yöntemlerle etkenin tespiti
- Kültür sonucu

Sample No	Time to blood positivity (hours)	Gram stain from positive bottle	Identification after solid agar incubation (18 hours)	
			Standart microbiology methods	/ Time to result (day)
1	11.5	Gr - bacilli	<i>Enterobacter spp.</i>	/ 4
2	8.2	Gr - bacilli	<i>E. coli</i>	/ 1
3	10.2	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
4	25.2	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
5	8.1	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
6	7.2	Gr + cocci	<i>Acinetobacter spp.</i>	/ 4
7	10.1	Gr + cocci	<i>Acinetobacter spp.</i>	/ 1
8	100.8	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
9	10.4	Gr - bacilli	<i>E. coli</i>	/ 1
10	12.1	Gr + cocci	<i>Enterococcus spp.</i>	/ 1
11	18.6	Gr + cocci	<i>Enterococcus spp.</i>	/ 2
12	14.1	Gr + cocci	<i>Micrococcus spp.</i>	/ 2
13	15.5	Gr - bacilli	<i>Morganella spp.</i>	/ 2
14	11.4	Gr + cocci	Contamination with skin flora	
15	38.6	Gr + cocci	Contamination with skin flora	
16	12.1	Gr - bacilli	<i>Enterobacter spp.</i>	/ 2
17	9.7	Gr + cocci	<i>Acinetobacter spp.</i>	/ 1
18	10.6	Gr - bacilli	<i>E. coli</i>	/ 1
19	71.1	Yeast	<i>Candida spp.</i>	/ 1
20	9.4	Gr - bacilli	<i>E. coli</i>	/ 1
21	12.4	Gr - bacilli	<i>Morganella spp.</i>	/ 1
22	8.2	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
23	11.1	Gr - bacilli	<i>E. coli</i> , ESBL (+)	/ 1
24	14.4	Gr - bacilli	<i>E. coli</i> , ESBL (+)	/ 1
25	10.1	Gr + cocci	<i>Acinetobacter spp.</i>	/ 1
26	17.4	Gr - bacilli	<i>P. aeruginosa</i>	/ 1
27	52.1	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
28	49.4	Gr - bacilli	<i>K. pneumoniae</i>	/ 1
29	8.8	Gr - bacilli	<i>E. coli</i> , ESBL (+), carbapenemase (+)	/ 2
30	12.2	Gr - bacilli	<i>E. coli</i> , ESBL (+), carbapenemase (+)	/ 1
31	10.4	Gr + cocci	<i>Acinetobacter spp.</i>	/ 1

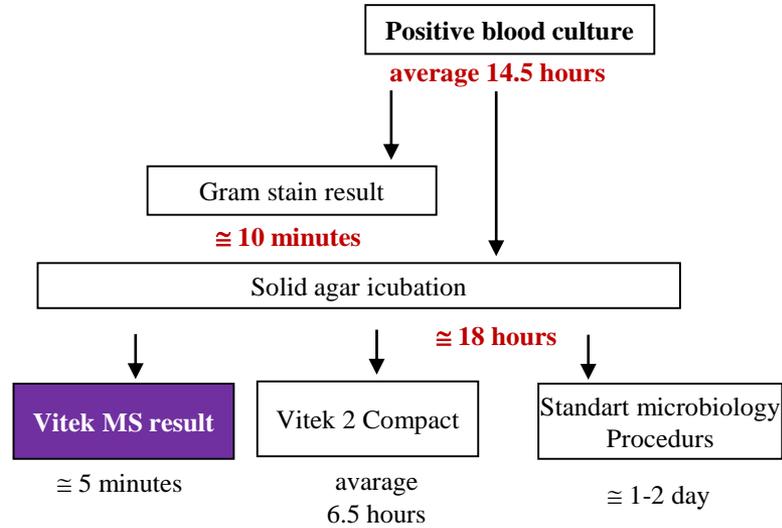
Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Kültür sonucu



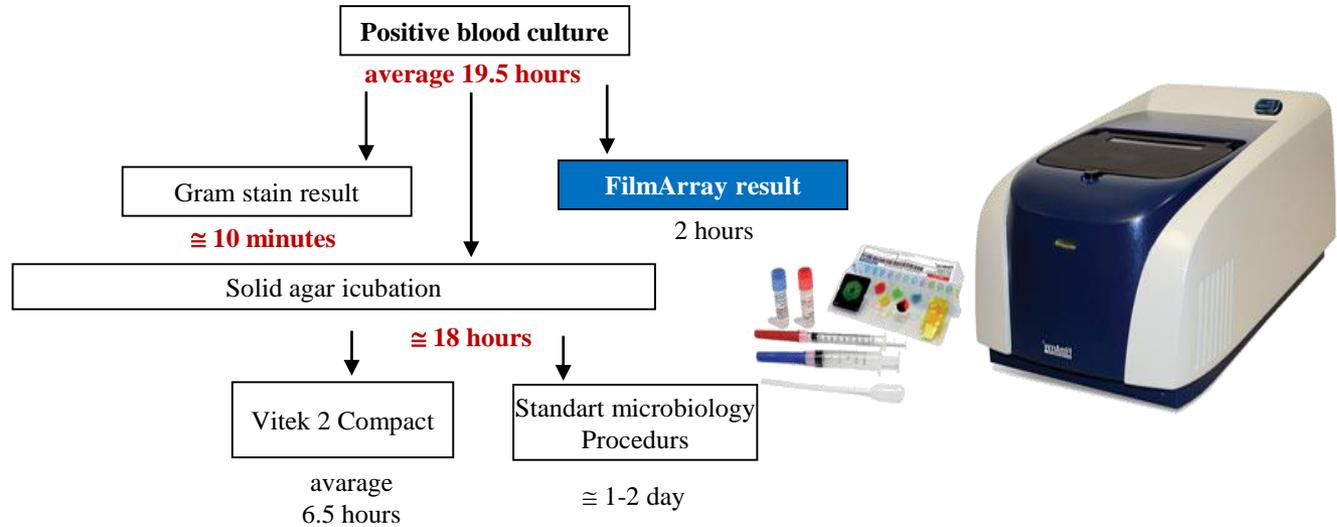
Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Kültür sonucu



Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti
 - Kültür sonucu



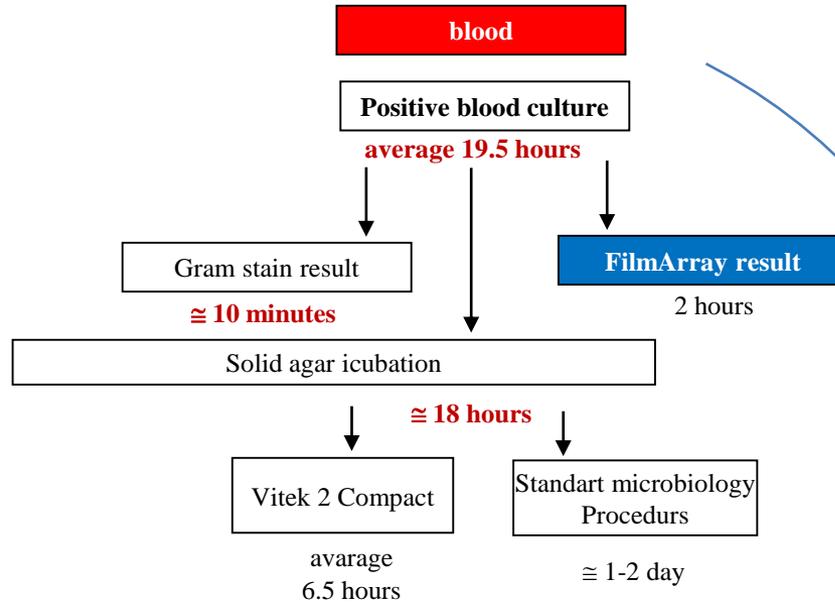
Enfeksiyon etkenin tespiti

- Moleküler yöntemlerle etkenin tespiti

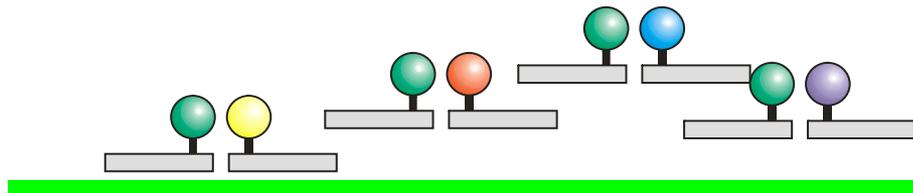
Sample No	Time to blood positivity (hours)	Gram stain from positive bottle	FilmArray Identification	Identification after solid agar incubation (18 hours)		
				Standart microbiology methods / Time to result (day)	Vitek 2 Compact / Time to result (hours)	Vitek MS
1	11.5	Gr - bacilli	<i>E. cloacae</i> complex	<i>Enterobacter</i> spp./ 4	<i>E. cloacae</i> complex / 5	<i>E. cloacae</i>
2	8.2	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> / 1	ND ^b	ND ^b
3	10.2	Gr - bacilli	<i>K. pneumoniae</i> , <i>P. aeruginosa</i>	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 4.15	<i>K. pneumoniae</i>
4	25.2	Gr - bacilli	<i>K. pneumoniae</i>	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 5	<i>K. pneumoniae</i>
5	8.1	Gr - bacilli	<i>K. pneumoniae</i> , <i>Streptococcus</i> spp	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 4.45	<i>K. pneumoniae</i>
6	7.2	Gr + cocci	<i>A. baumannii</i>	<i>Acinetobacter</i> spp./ 4	<i>A. baumannii</i> complex / 6	<i>A. baumannii</i> complex
7	10.1	Gr + cocci	<i>A. baumannii</i>	<i>Acinetobacter</i> spp./ 1	<i>A. baumannii</i> complex / 6	<i>A. baumannii</i> complex
8	100.8	Gr - bacilli	<i>K. pneumoniae</i>	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 6	<i>K. pneumoniae</i>
9	10.4	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> / 1	ND ^b	ND ^b
10	12.1	Gr + cocci	<i>Enterococcus</i> spp., Van A/B (-)	<i>Enterococcus</i> spp./ 1	<i>E. faecium</i> / 6	<i>E. faecium</i>
11	18.6	Gr + cocci	<i>Enterococcus</i> spp., Van A/B (-)	<i>Enterococcus</i> spp./ 2	<i>E. faecium</i> / 6	<i>E. faecium</i>
12	14.1	Gr + cocci	Not detected	<i>Micrococcus</i> spp./ 2	<i>E. cloacae</i> / 5	<i>E. cloacae</i>
13	15.5	Gr - bacilli	<i>Staphylococcus</i> spp., <i>MecA</i> (-)	<i>Morganella</i> spp./ 2	<i>Morganella morganii</i> / 5.45	<i>Morganella morganii</i>
14	11.4	Gr + cocci	<i>Enterococcus</i> spp., Van A/B (-) <i>Staphylococcus</i> spp., <i>MecA</i> (-)	Contamination with skin flora	ND ^c	ND ^c
15	38.6	Gr + cocci	<i>A. baumannii</i> <i>Enterococcus</i> spp., Van A/B (-)	Contamination with skin flora	ND ^c	ND ^c
16	12.1	Gr - bacilli	<i>E. cloacae</i> complex, KPC (-)	<i>Enterobacter</i> spp., carbapenemase (+) / 2	<i>E. cloacae</i> complex / 5.15	<i>E. cloacae</i>
17	9.7	Gr + cocci	<i>A. baumannii</i>	<i>Acinetobacter</i> spp./ 1	<i>A. baumannii</i> complex / 6	<i>A. baumannii</i> complex
18	10.6	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> , ESBL (+)/ 1	ND ^b	ND ^b
19	71.1	Yeast	<i>C. tropicalis</i>	<i>Candida</i> spp./ 1	<i>C. tropicalis</i> / 18.15	<i>C. kefyr</i> ^d
20	9.4	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> / 1	ND ^b	ND ^b
21	12.4	Gr - bacilli	Not detected	<i>Morganella</i> spp./ 1	<i>Morganella morganii</i> / 5.45	<i>Morganella morganii</i>
22	8.2	Gr - bacilli	<i>K. pneumoniae</i> KPC(-), <i>Enterococcus</i> Van A/B(-)	<i>K. pneumoniae</i> , carbapenemase (+) / 1	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
23	11.1	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> , ESBL (+), carbapenemase (+) / 1	ND ^b	ND ^b
24	14.4	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> , ESBL (+), carbapenemase (+) / 1	ND ^b	ND ^b
25	10.1	Gr + cocci	<i>A. baumannii</i>	<i>Acinetobacter</i> spp./ 1	<i>A. baumannii</i> complex / 8	<i>A. baumannii</i> complex
26	17.4	Gr - bacilli	<i>P. aeruginosa</i>	<i>P. aeruginosa</i> / 1	<i>P. aeruginosa</i> / 4.45	<i>P. aeruginosa</i>
27	52.1	Gr - bacilli	<i>K. pneumoniae</i> , KPC (-)	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 4.45	<i>K. pneumoniae</i>
28	49.4	Gr - bacilli	<i>K. pneumoniae</i> KPC (-)	<i>K. pneumoniae</i> / 1	<i>K. pneumoniae</i> / 4	<i>K. pneumoniae</i>
29	8.8	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> , ESBL (+), carbapenemase (+) / 2	ND ^b	ND ^b
30	12.2	Gr - bacilli	<i>E. coli</i> , KPC (-)	<i>E. coli</i> , ESBL (+), carbapenemase (+) / 1	ND ^b	ND ^b
31	10.4	Gr + cocci	<i>A. baumannii</i>	<i>Acinetobacter</i> spp./ 1	<i>A. baumannii</i> complex / 6.5	<i>A. baumannii</i> complex

Enfeksiyon etkeninin tespiti

- Moleküler yöntemlerle etkenin tespiti



The LightCycler® SeptiFast Test



Enfeksiyon etkenin tespiti

- Moleküler yöntemlerle etkenin tespiti

Curiosity Diagnostics

PCR|ONE For Investors About Us

Help when you need it most

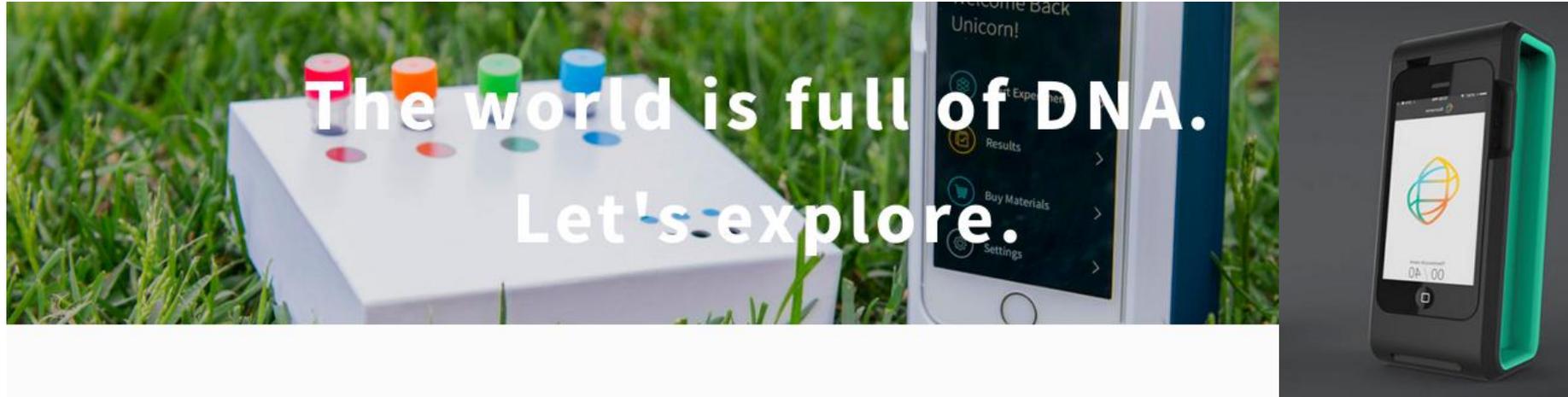
(I)
COLLECT SAMPLE



SEPSIS
GOLDEN HOUR

Saving patients with systemic infections requires rapid administration of drugs. PCR|ONE system allows drugs to be specifically targeted for optimal treatment.

Enfeksiyon etkeninin tespiti

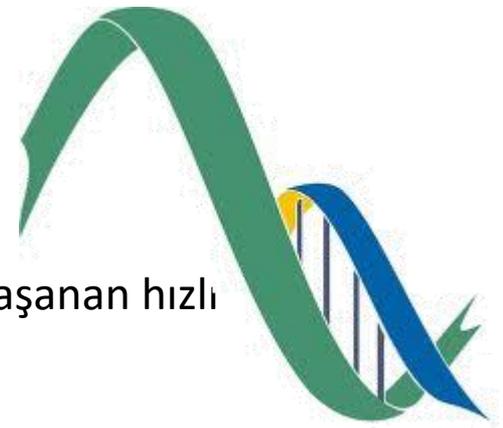


We are Biomeme:
A smartphone-based DNA detection platform.
No lab necessary.

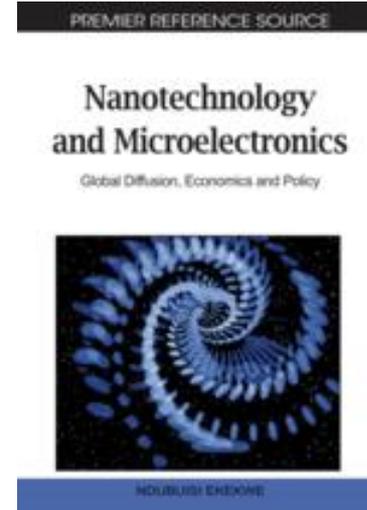
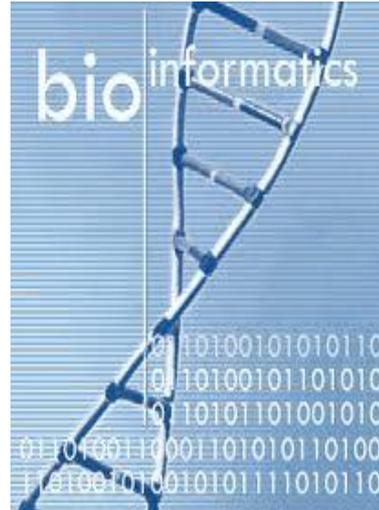
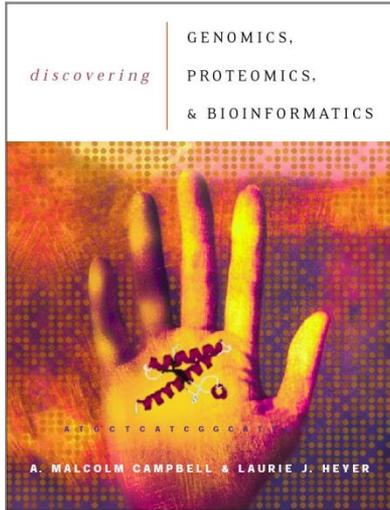
Enfeksiyon etkeninin tespiti



Biomeme Sample Prep for Children



- Genomik, biyoinformatik ve mikroelektronik alanında yaşanan hızlı gelişmelerin en göze çarpan sonuçları,
 - **Biyosensörlerdir ve**
 - **DNA mikroçip teknolojileri**



Biyosensörler



Biyosensörler



BIO-X *Solutions for Hospital-Acquired Infections*



FOTO: STAFFAN CLAESSEON

BIO-X® offers a structured approach to partnering with innovators from academia, clinics and biotech SME's to develop proof of concept, proof of mechanisms or proof of hypothesis for new life science products and services. The BIO-X program offers selected project teams tailor-made process support and financing, up to 1 million SEK per year for up to two

years. We are currently looking for healthcare for projects seeking to develop diagnostics and other solutions to combat hospital-acquired infections.

The recent BIO-X Call for proposals for new solutions for fighting hospital acquired infections generated interest in academic research, clinics as well as s

- **Rapid and sensitive diagnostic bench-top system for detection of hospital-acquired infections**
A fully automated microfluidic benchtop system for rapid, sensitive and decentralized detection of hospital-acquired infections, based on magnetic bioassays.
- **Antibacterial polymers for prevention of surgical site and wound infections**
Antibacterial and biocompatible polymers for prevention of surgical site infections and wound infections.
- **Sampling device for simultaneous transportation and enrichment of multidrug-resistant bacteria samples**
A sampling device for simultaneous transportation and broth enrichment of multidrug-resistant bacteria to increase sensitivity and shorten the screening process.
- **Cranioplasty implant for large skull defects limiting hospital-acquired infections**
A bio-ceramic implant for use in cranioplasty of large skull defects limiting bacterial infections related to replacement of the skull bone.

- **Biosensors for early diagnostics of hospital-acquired infections**
An optical biosensor platform technology for early diagnosis of infectious agents on a multitude of surfaces, eg on medical devices, in patients' wounds, etc.

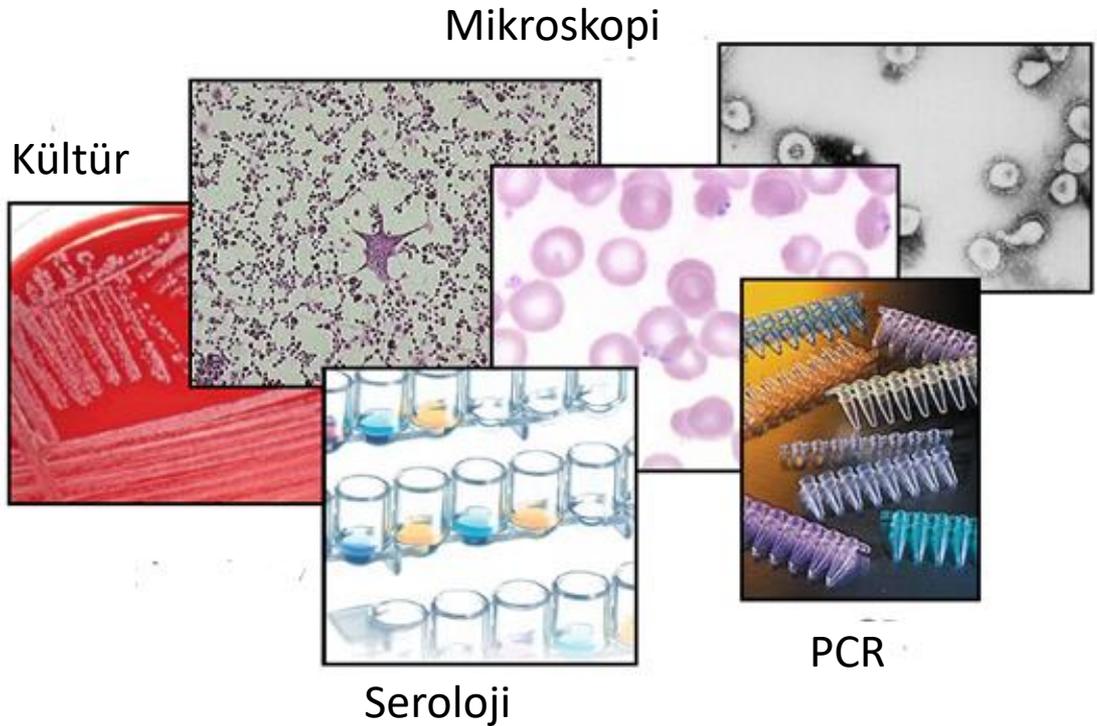
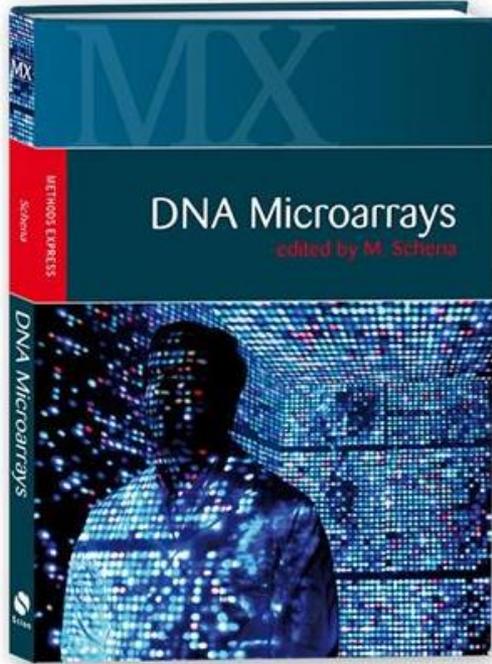
Antimicrobial surface technology for covalently coating medical grade materials for reducing hospital acquired infections; in this instance, single use silicon based devices used in ventilatory support.

- **Antifungal coating for medical devices**
Antifungal coating technology to prevent in-growth of fungal hyphae and prevent biofilm formation on medical devices.
- **Ultrafast lab-on-a-chip system for microbial detection**
An antibody-based ultrafast, sensitive, lab-on-a-chip system for microbial

DNA Mikroçip Teknolojileri

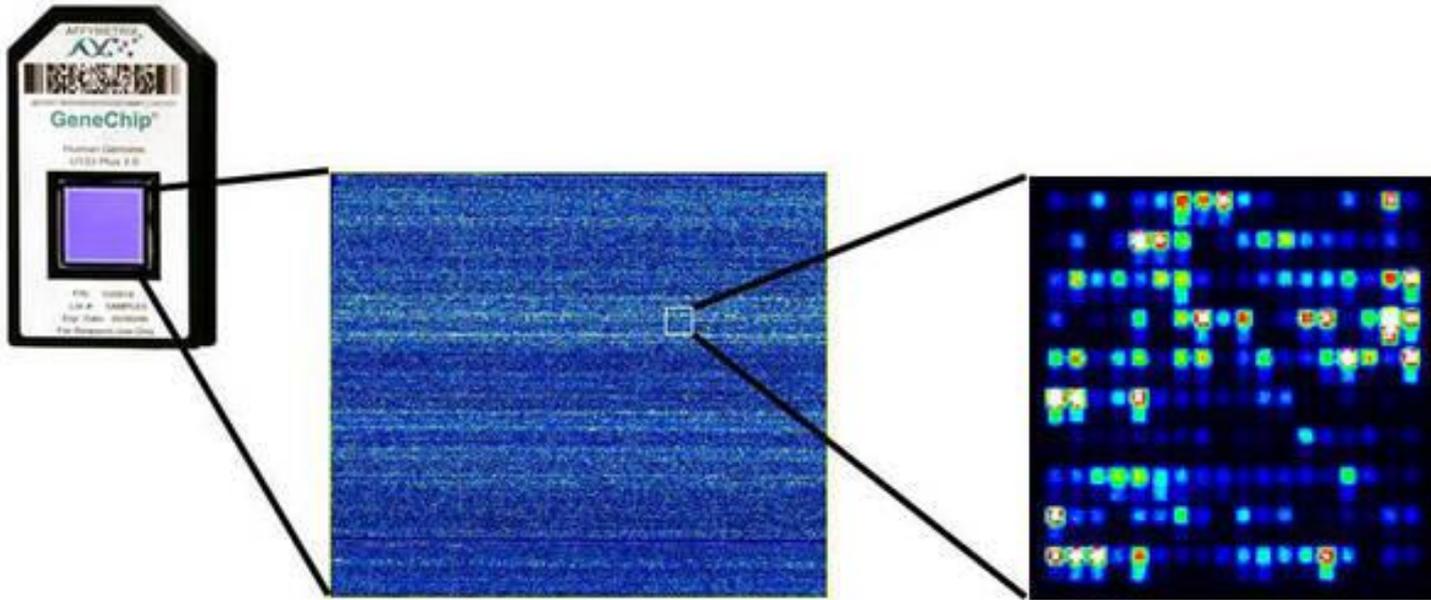
- DNA mikroçip teknolojileri, çevresel ve klinik örneklerden **mikroorganizmaların tanısı için** giderek artan oranda kullanılmaktadır.

Geleneksel Mikrobiyolojik Tanı Yöntemleri



DNA Mikroçip Teknolojileri

- PCR ile elde edilen floresanla işaretli ampliconların çok sayıda farklı **oligonükleotid prob** içeren katı yüzeylerde, kendisine uyan proba hibridize olması temeline dayanmaktadır.



DNA Mikroçip Teknolojileri

- **Virochip** 1500 virüse ait 36.000 prob içermektedir.

Using a Pan-Viral Microarray Assay (Virochip) to Screen Clinical Samples for Viral Pathogens

Eunice C. Chen¹, Steve A. Miller¹, Joseph L. DeRisi^{1,2}, Charles Y. Chiu^{1,2}

¹Department of Laboratory Medicine, University of California, San Francisco

²Division of Infectious Diseases, University of California, San Francisco

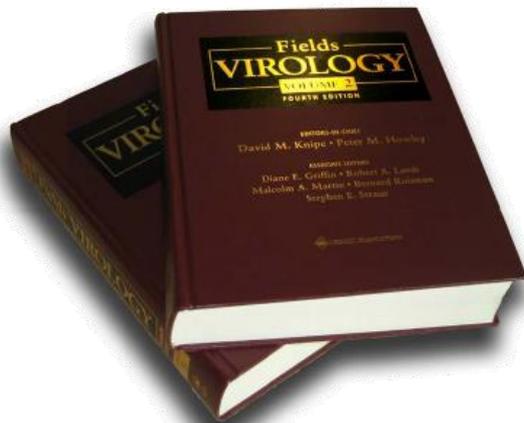
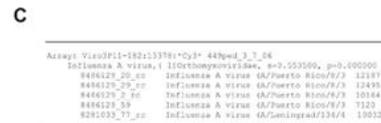
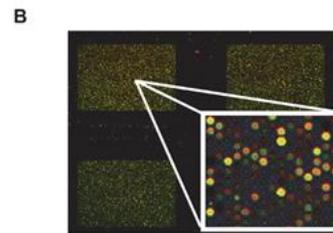


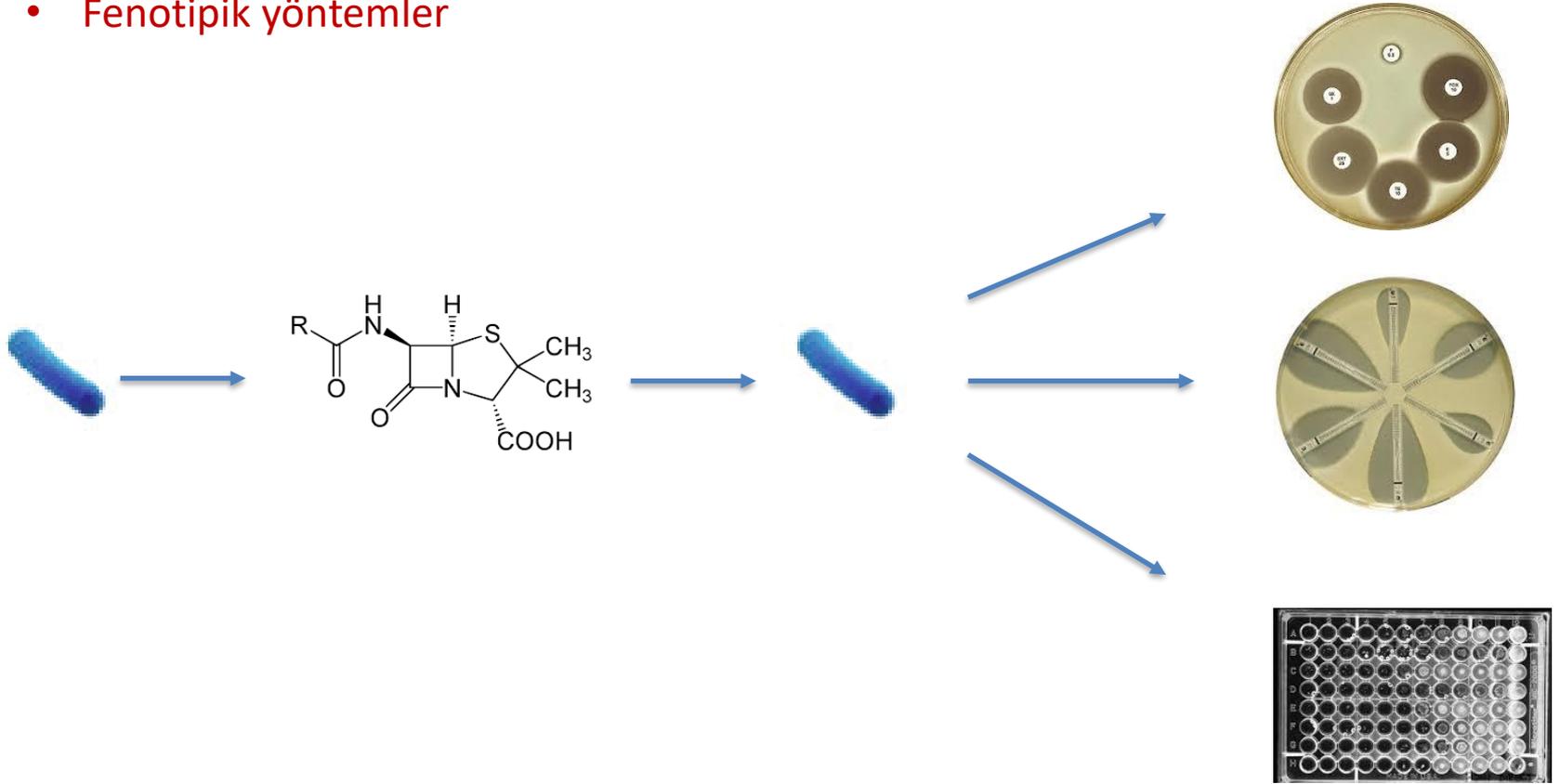
Figure 2. Steps in the Virochip assay. After amplification by random PCR, a smear of 200 - 1000 bp can be visualized by gel electrophoresis (A). (B) Three Virochip microarrays out of the 8 arrays / glass slide are shown, with a small region of one microarray blown-up in the inset on the bottom right corner. (C) Automated microarray viral analysis using E-Predict revealing the presence of influenza A virus in the clinical sample.



Antibiyotik Direncini Tespit Etmek

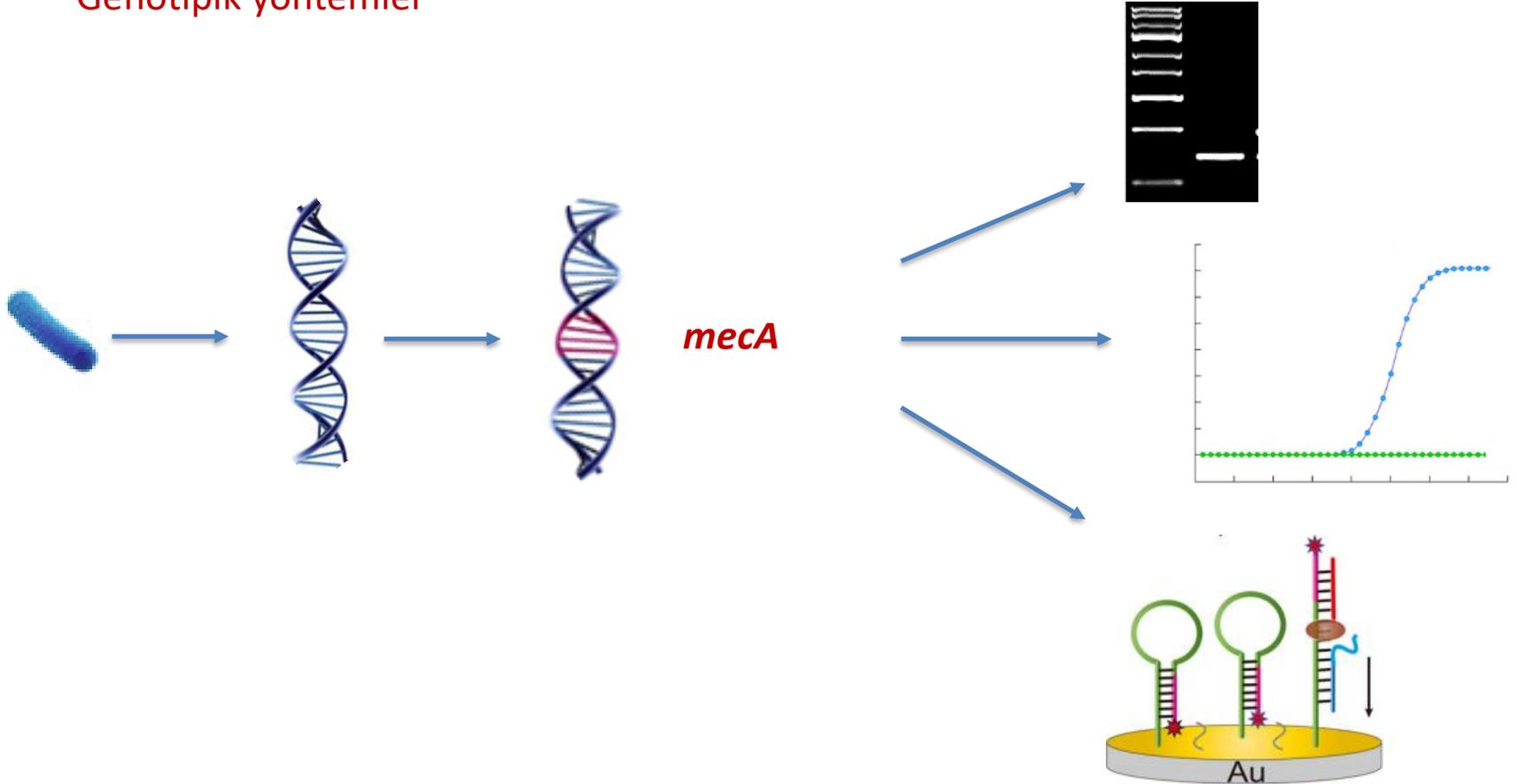
- Antibiyotik direncini tespit etmek için genel olarak **iki yaklaşım** mevcuttur.

- Fenotipik yöntemler



Antibiyotik Direncini Tespit Etmek

- Antibiyotik direncini tespit etmek için genel olarak iki yaklaşım mevcuttur.
 - Genotipik yöntemler



Antibiyotik Direncini Tespit Etmek

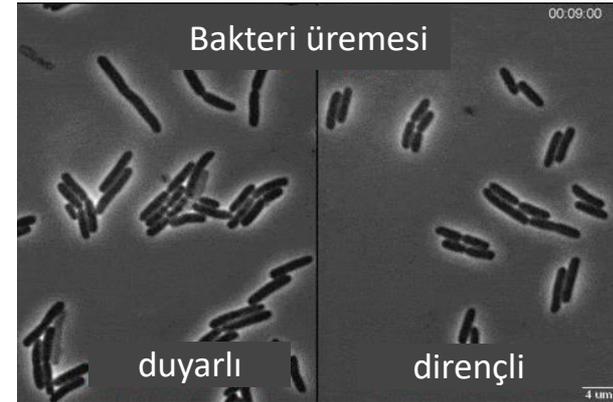
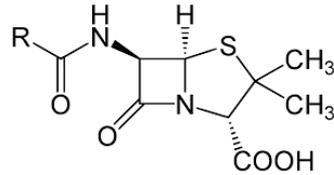
- Antibiyotik duyarlılığı ancak fenotipik yöntemlerle belirlenebilir !

mikroorganizma ile antibiyotik karşılaştırılmadan duyarlılık sonucundan bahsedilemez

S. aureus

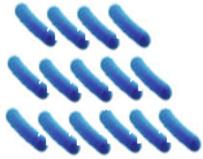


Penisilin

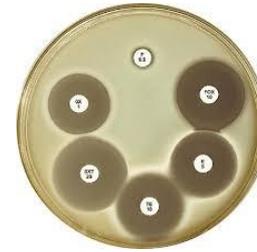


Antibiyotik Direncini Tespit Etmek

- Yeni fikirler, yeni sistemler
 - Fenotipik yöntemlerde **üremenin/inhibisyonun** gösterilmesi zaman alıcı.



18-24 saat



16-18 saat



8-10 saat



6-8 saat

Antibiyotik Direncini Tespit Etmek

- Fenotipik testlerdeki gelişmeler;

mikroorganizma üremesinin ya da üremenin inhibisyonunun hızla tespit edilmesi

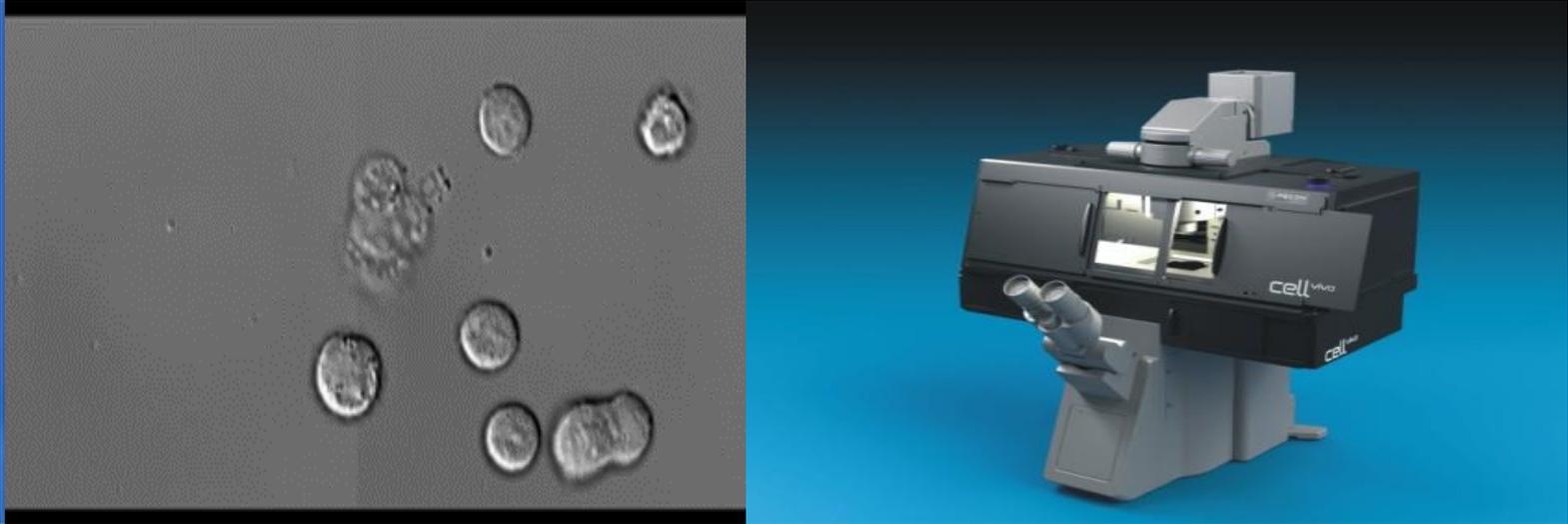
temeline dayanmaktadır.

- Kolorimetrik
- Biolüminesans
- Kemilüminesans
- Nefelometrik
- Spektrometrik cihazlar
- Zaman-atlamalı mikroskop
- Raman-spektrometresi
- Atomik güç mikroskobu

Antibiyotik Direncini Tespit Etmek

Zaman Atlamalı Mikroskopik Yöntemler

- Hücrelerin canlı kalarak çoğalabildikleri bir ortamda, **belirli zamana aralıklarında** çekim yapabilen mikroskoplar.



Antibiyotik Direncini Tespit Etmek

Zaman Atlamalı Mikroskopik Yöntemler

- Mikroorganizmalar üzerine antibiyotik etkisi **6 – 30 dakikada** tespit edilebiliyor.



Real-Time Optical Antimicrobial Susceptibility Testing

Marlene Fredborg,^a Klaus R. Andersen,^b Erik Jørgensen,^b Aida Droce,^{a,c} Tom Olesen,^b Bent B. Jensen,^a Flemming S. Rosenvinge,^d Teis E. Sondergaard^{a,c}

Department of Animal Science, Faculty of Science and Technology, Aarhus University, Tjele, Denmark^a; Unisensor Ltd., Allerød, Denmark^b; Department of Biotechnology, Chemistry and Environmental Engineering, Aalborg University, Aalborg, Denmark^c; Odense University Hospital, Department of Clinical Microbiology, Odense, Denmark^d

Rapid antibiotic susceptibility testing is in high demand in health care fields as antimicrobial-resistant bacterial strains emerge and spread. Here, we describe an optical screening system (oCelloScope) which, based on time-lapse imaging of 96 bacteria-antibiotic combinations at a time, introduces real-time detection of bacterial growth and antimicrobial susceptibility with imaging material to support the automatically generated graphs. Automated antibiotic susceptibility tests of a monoculture showed statistically significant antibiotic effects within 6 min and within 30 min in complex samples from pigs suffering from catheter-associated urinary tract infections. The oCelloScope system provides a fast high-throughput screening method for detecting bacterial susceptibility that might entail an earlier diagnosis and introduction of appropriate targeted therapy and thus combat the threat from multidrug-resistant pathogenic bacteria. The oCelloScope system can be employed for a broad range of applications within bacteriology and might present new vistas as a point-of-care instrument in clinical and veterinary settings.

Antibiyotik Direncini Tespit Etmek

Zaman Atlamalı Mikroskobik Yöntemler

Fredborg et al.



FIG 1 The oCelloScope detection system. (a) P detection principle. A volume of 50 µl of a 96-dimensional (2D) picture. (c) 2D picture of *S. al* microscope.

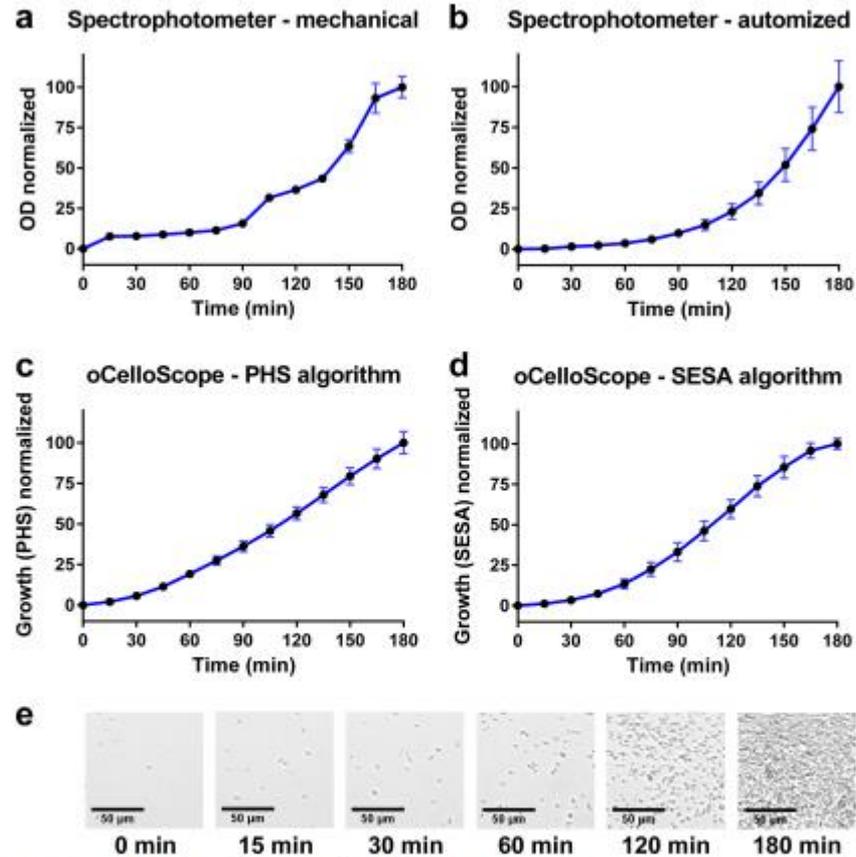
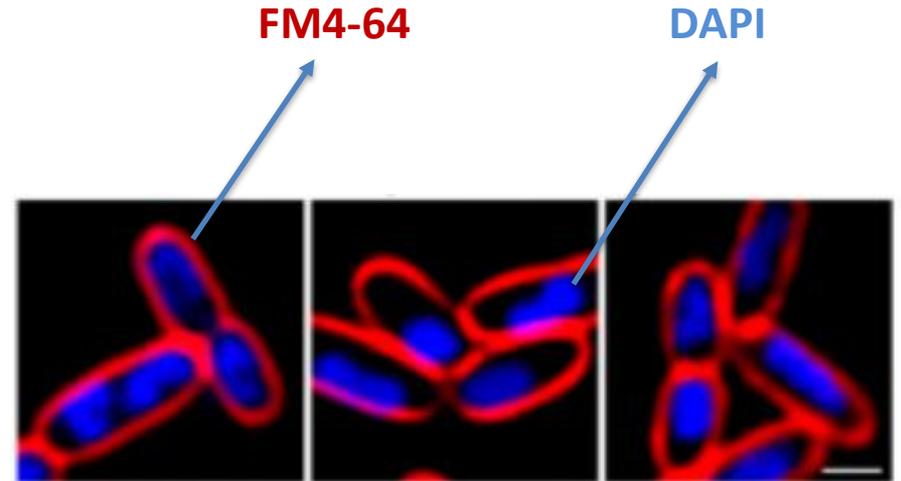
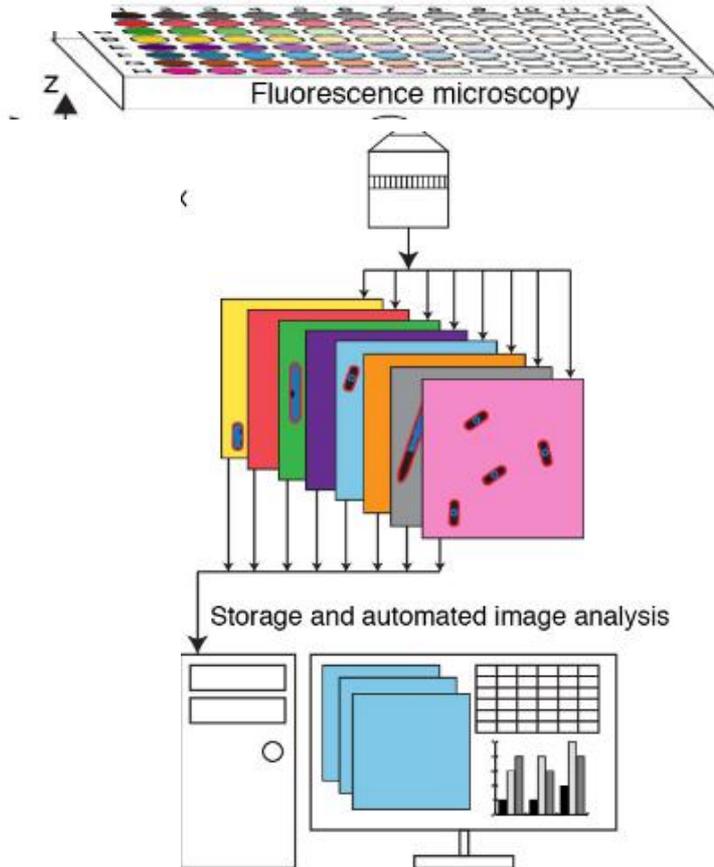


FIG 2 Bacterial growth of *S. aureus* assessed by the oCelloScope and traditional OD measurements. (a) Growth was measured by optical density in a standard laboratory spectrophotometer with one cuvette. The absorbance was measured at 600 nm. (b) Growth was measured by optical density (absorbance, 655 nm) using a standard laboratory plate reader with a 96-well plate. (c) Growth was measured by optical density using the oCelloScope pixel histogram summation (PHS) algorithm. (d) Growth was measured by the oCelloScope segmentation and extraction of surface area (SESA) algorithm. (e) Pictures taken by the oCelloScope showing bacterial growth to different time points. All experiments were done as eight replicates, and standard derivations are shown as error bars on the curves. Scale bar, 50 µm.

Antibiyotik Direncini Tespit Etmek

Floresans Mikroskopik Yöntemler

- *Bacterial cytological profiling (BCP)*, bakterilerin morfolojik analizi ile antibiyotik duyarlılık



Antibiyotik Direncini Tespit Etmek

Florens Mikroskopik Yöntemler

- *Bacterial cytological profiling*, %100 doğrulukla 1-2 saat için metisilin 30 dakikada daptomisin direnci tespiti



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Research Paper

Bacterial Cytological Profiling (BCP) as a Rapid and Accurate Antimicrobial Susceptibility Testing Method for *Staphylococcus aureus*



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ARTICLE INFO

Article history:

Received 30 November 2015

Received in revised form 7 January 2016

Accepted 15 January 2016

Available online 18 January 2016

Keywords:

Antibiotic resistance

Susceptibility tests

Staphylococcus aureus

Multidrug resistant bacteria

ABSTRACT

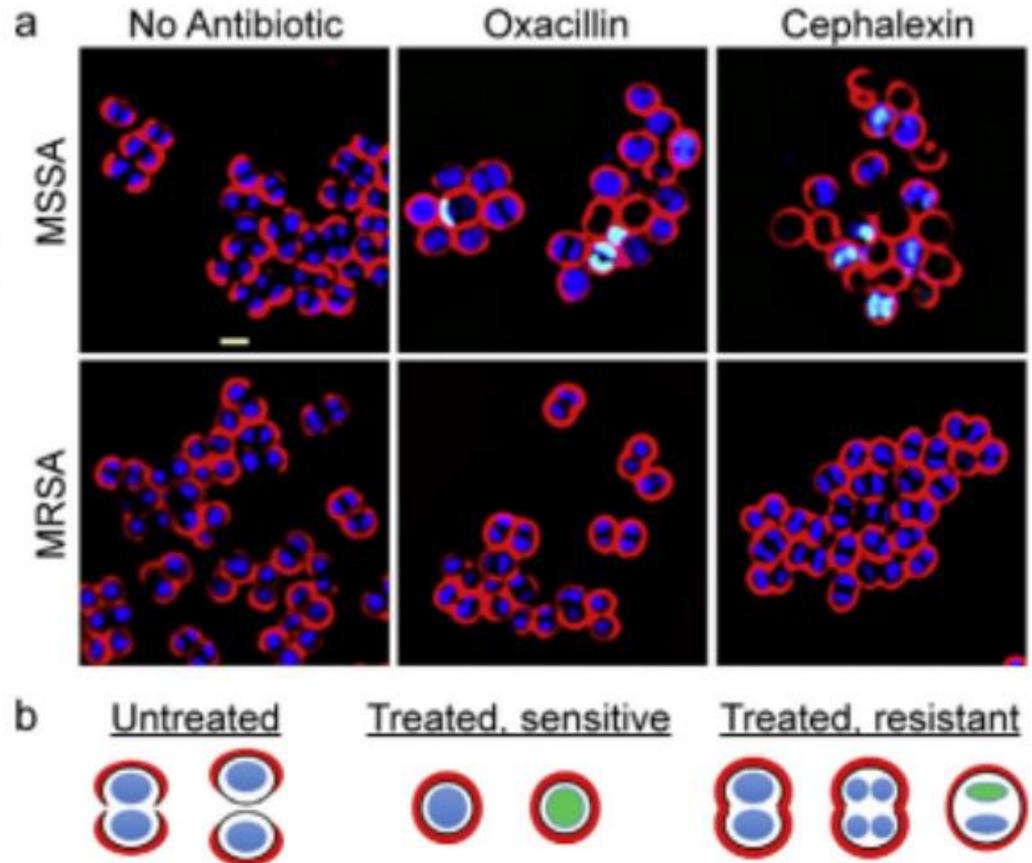
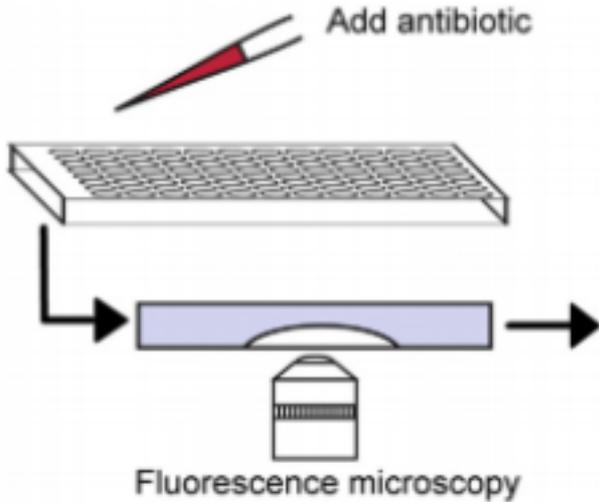
Successful treatment of bacterial infections requires the timely administration of appropriate antimicrobial therapy. The failure to initiate the correct therapy in a timely fashion results in poor clinical outcomes, longer hospital stays, and higher medical costs. Current approaches to antibiotic susceptibility testing of cultured pathogens have key limitations ranging from long run times to dependence on prior knowledge of genetic mechanisms of resistance. We have developed a rapid antimicrobial susceptibility assay for *Staphylococcus aureus* based on bacterial cytological profiling (BCP), which uses quantitative fluorescence microscopy to measure antibiotic induced changes in cellular architecture. BCP discriminated between methicillin-susceptible (MSSA) and -resistant (MRSA) clinical isolates of *S. aureus* ($n = 71$) within 1–2 h with 100% accuracy. Similarly, BCP correctly distinguished daptomycin susceptible (DS) from daptomycin non-susceptible (DNS) *S. aureus* strains ($n = 20$) within 30 min. Among MRSA isolates, BCP further identified two classes of strains that differ in their susceptibility to specific combinations of beta-lactam antibiotics. BCP provides a rapid and flexible alternative to gene-based susceptibility testing methods for *S. aureus*, and should be readily adaptable to different antibiotics and bacterial species as new mechanisms of resistance or multidrug-resistant pathogens evolve and appear in mainstream clinical practice.

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Antibiyotik Direncini Tespit Etmek

Optikal Difüzetri

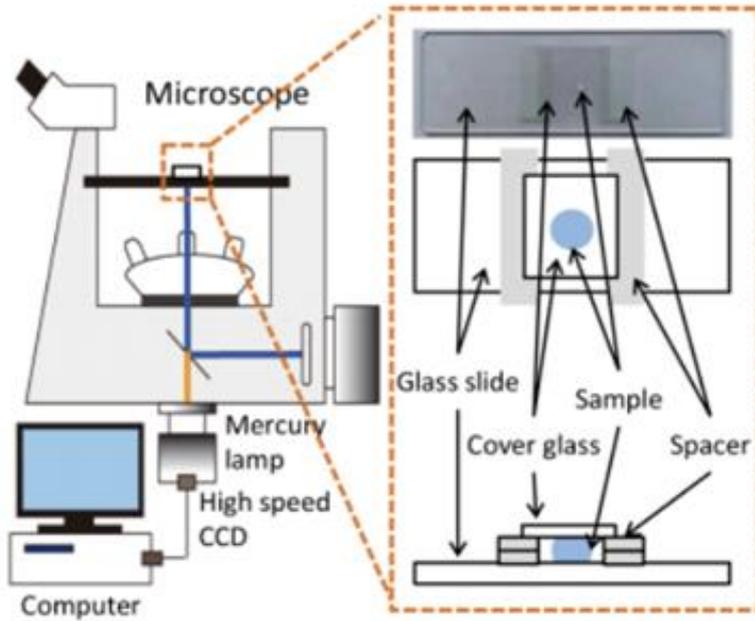
- SYTOX, DAPI, WGA, FM-64 gibi floresan boyalar



Antibiyotik Direncini Tespit Etmek

Optikal Difüzetri

- Parçacıkların **difüzyon katsayılarını** ölçmek için tasarlanmış bir sistemdir.



Antibiyotik Direncini Tespit Etmek

Optikal Difüzometri

- **Optikal difüsometri ve bead-based immunoassays** yöntemlerinin kombine edildiği yöntemde, *P. aeruginosa*'da gentamisin direnci iki saat içerisinde tespit edilebilmiştir.



RESEARCH ARTICLE

Rapid Bead-Based Antimicrobial Susceptibility Testing by Optical Diffusometry

Chih-Yao Chung¹, Jhih-Cheng Wang^{1,2}, Han-Sheng Chuang^{1,3*}

1 Department of Biomedical Engineering, National Cheng Kung University, Tainan, Taiwan, **2** Division of Urology, Department of Surgery, Chi Mei Medical Center, Tainan, Taiwan, **3** Medical Device Innovation Center, National Cheng Kung University, Tainan, Taiwan

* oswaldchuang@mail.ncku.edu.tw



Abstract

This study combined optical diffusometry and bead-based immunoassays to develop a novel technique for quantifying the growth of specific microorganisms and achieving rapid AST. Diffusivity rises when live bacteria attach to particles, resulting in additional energy from motile microorganisms. However, when UV-sterilized (dead) bacteria attach to particles, diffusivity declines. The experimental data are consistent with the theoretical model predicted according to the equivalent volume diameter. Using this diffusometric platform, the susceptibility of *Pseudomonas aeruginosa* to the antibiotic gentamicin was tested. The result suggests that the proliferation of bacteria is effectively controlled by gentamicin. This study demonstrated a sensitive (one bacterium on single particles) and time-saving (within 2 h) platform with a small sample volume (~0.5 μL) and a low initial bacteria count (50 CFU per droplet ~ 10^5 CFU/mL) for quantifying the growth of microorganisms depending on Brownian motion. The technique can be applied further to other bacterial strains and increase the success of treatments against infectious diseases in the near future.

OPEN ACCESS

Citation: Chung C-Y, Wang J-C, Chuang H-S (2016) Rapid Bead-Based Antimicrobial Susceptibility Testing by Optical Diffusometry. PLoS ONE 11(2): e0148864. doi:10.1371/journal.pone.0148864

Editor: Bing-Yang Cao, Tsinghua University, CHINA

Received: October 27, 2015

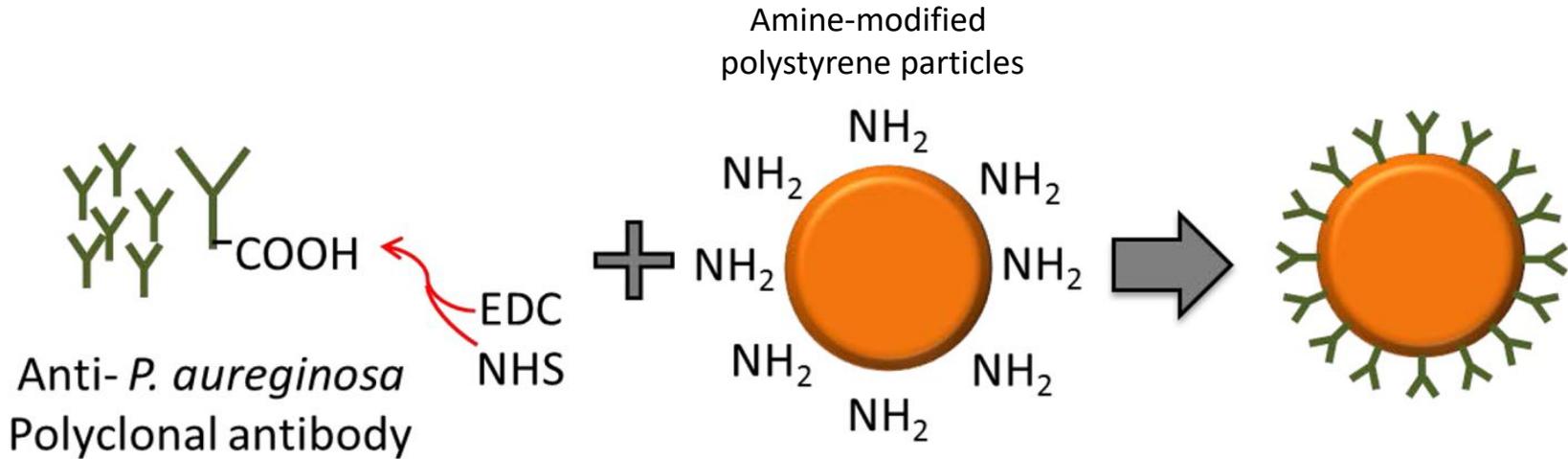
Accepted: January 25, 2016

Published: February 10, 2016

Antibiyotik Direncini Tespit Etmek

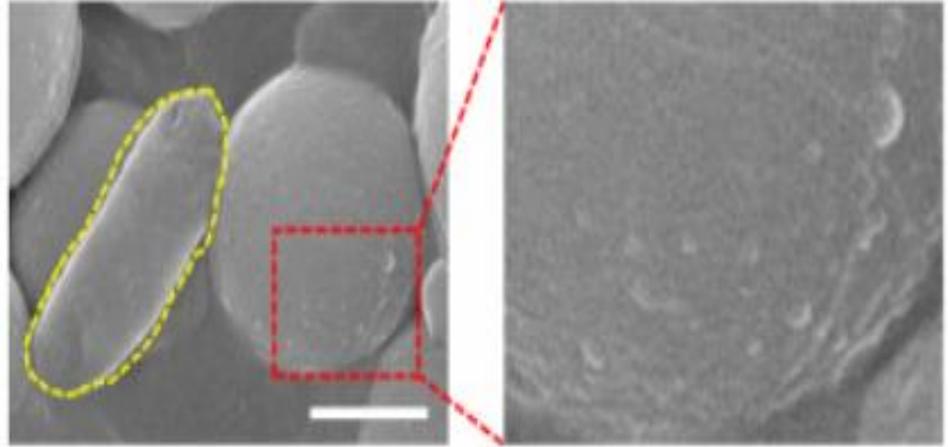
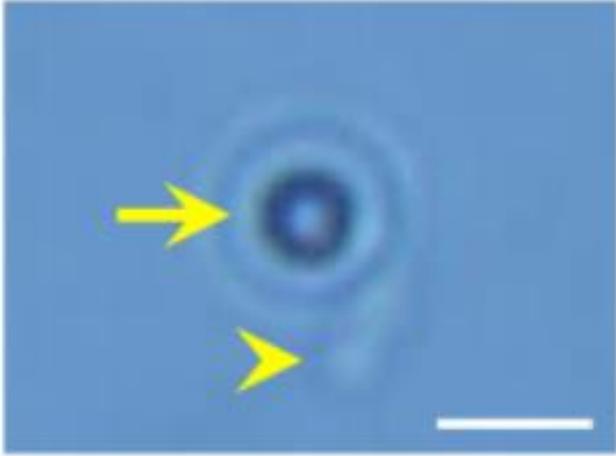
Optikal Difüzometri

- Antikorun aktive edilmesinde **EDC-NHS kimyasalları** kullanılarak antikorun polystyrene matrikse bağlanması.



Optikal Difüzometri

- Boncuklar üzerinde yakalanmış *P. aeruginosa*.



Optikal Difüzometri

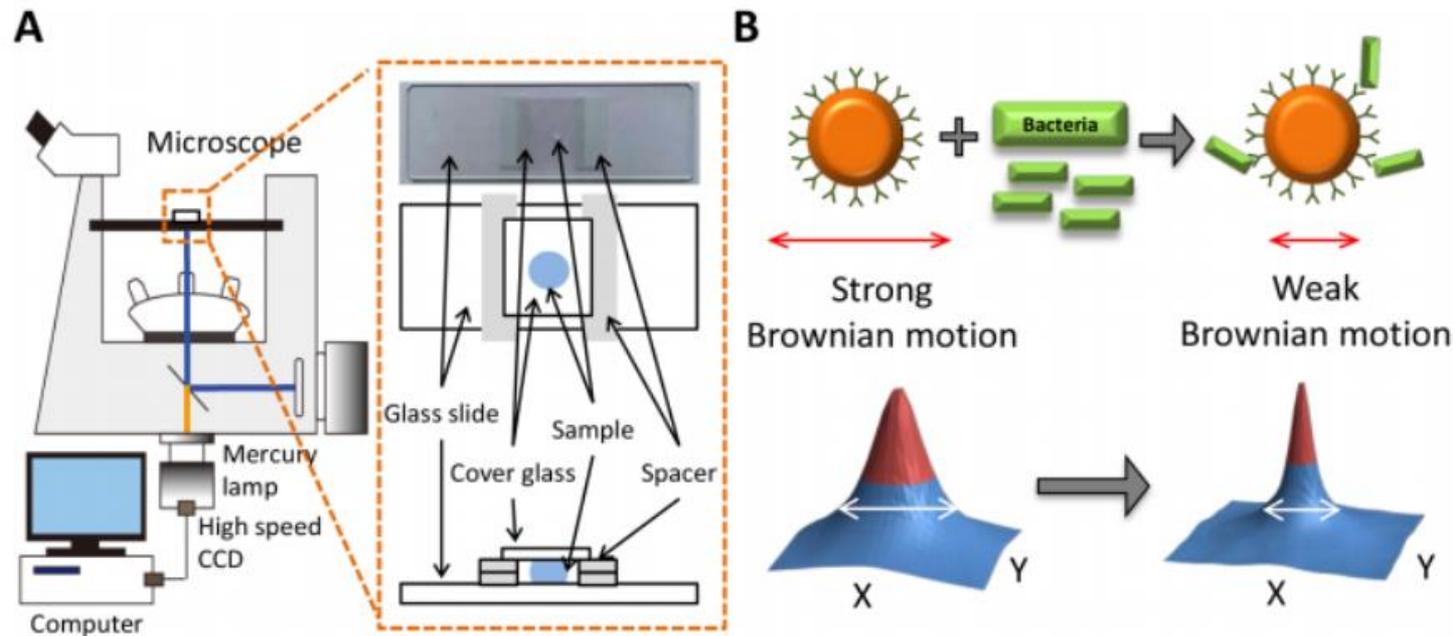
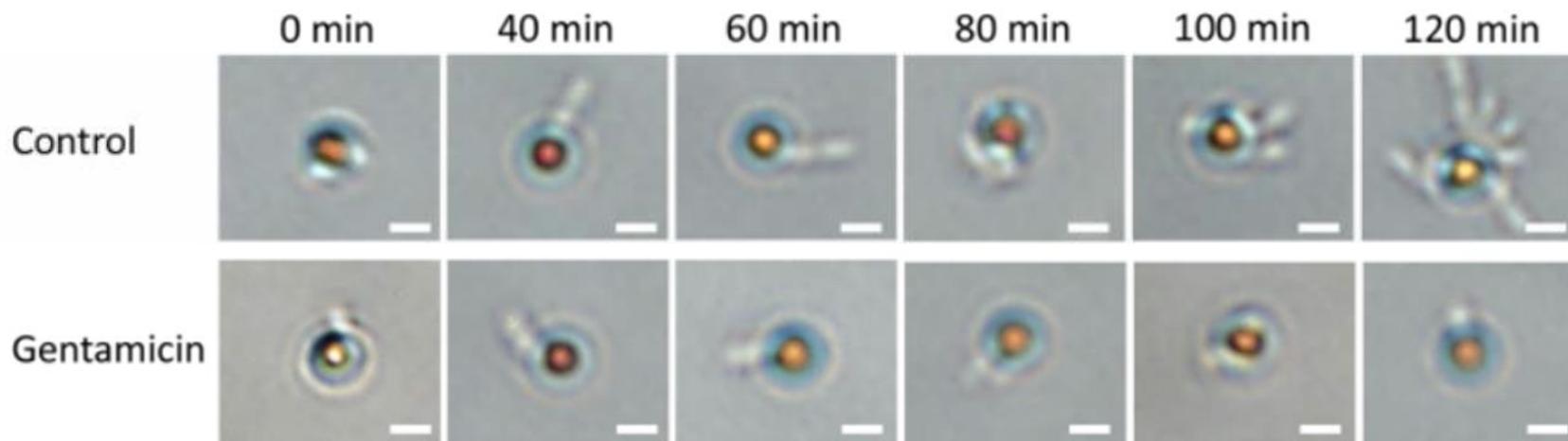
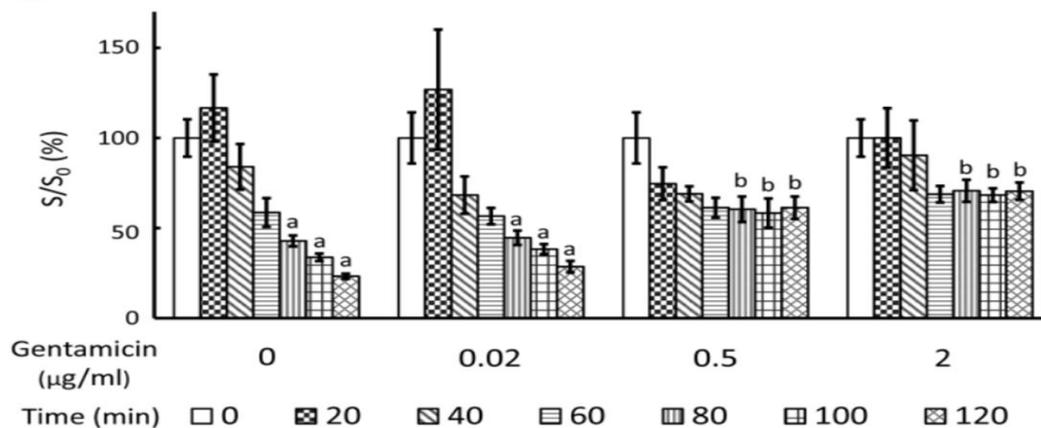


Fig 1. The optical diffusometric platform. (A) Schematic of the optical diffusometry. (B) The relationship of Brownian motion and the particle size change due to the bacterium-particle binding. The corresponding diffusivity values are derived from the cross-correlation algorithm. A large particle diameter results in a narrow correlation peak.

Optikal Difüzmometri



B



Ticari sistem- Accelerate Diagnostics Pheno System

- Accelerate Diagnostics; direkt örnekten hızlı tanımlama ve duyarlılık



Accelerate Diagnostics - Pheno System

- Örnekteki bakteriyi saflaştırma ve çoğaltmak

1 saat

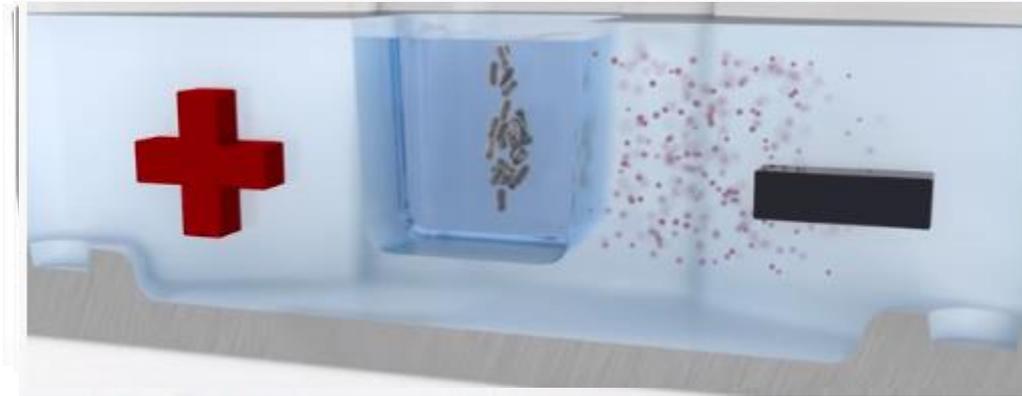


8-48 saat



Accelerate Diagnostics - Pheno System

- Bakteri hücrelerin örnek muayene maddesinden saflaştırılması için **elektroforez ve filtreleme sistemi** (elektrofiltrasyon).



Accelerate Diagnostics - Pheno System

- Mikroorganizmanın tanımlanması

1 saat



8-48 saat

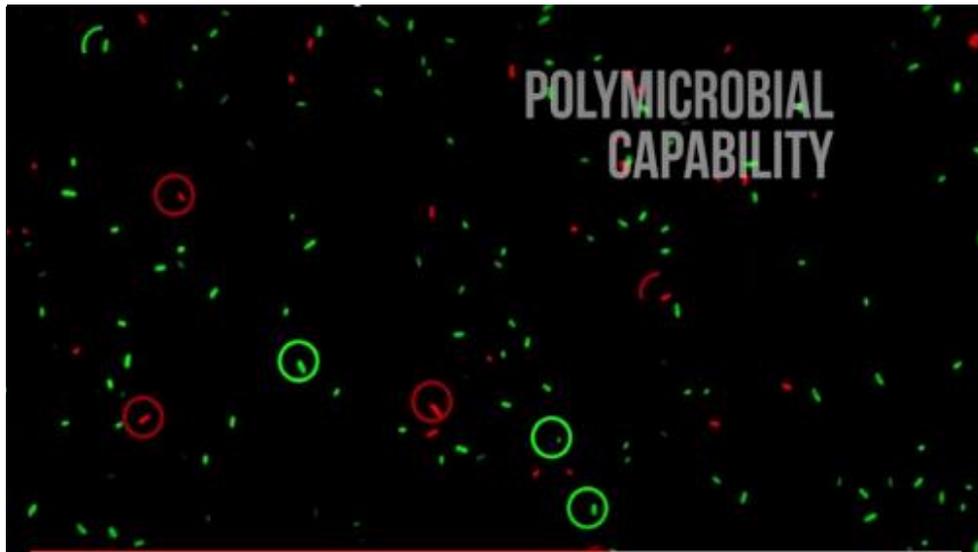


Accelerate Diagnostics - Pheno System

- Multiplexed Fluorescence in situ Hybridization (FISH)

Accelerate Diagnostics - Pheno System

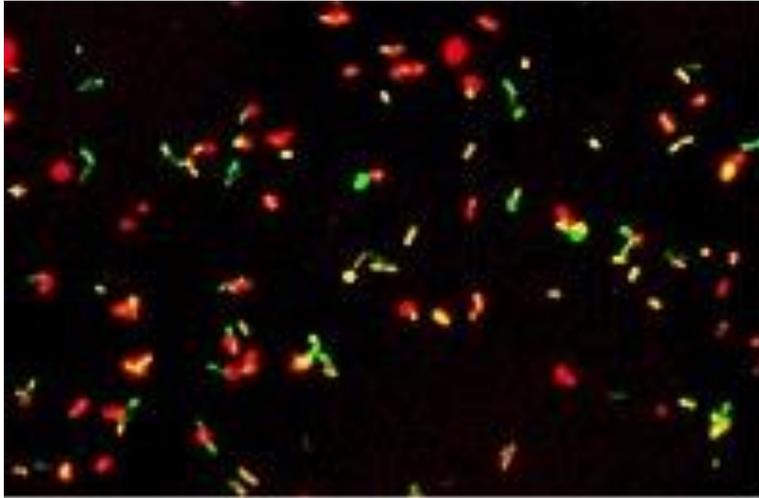
- *Multiplexed Fluorescence in situ Hybridization (FISH)*



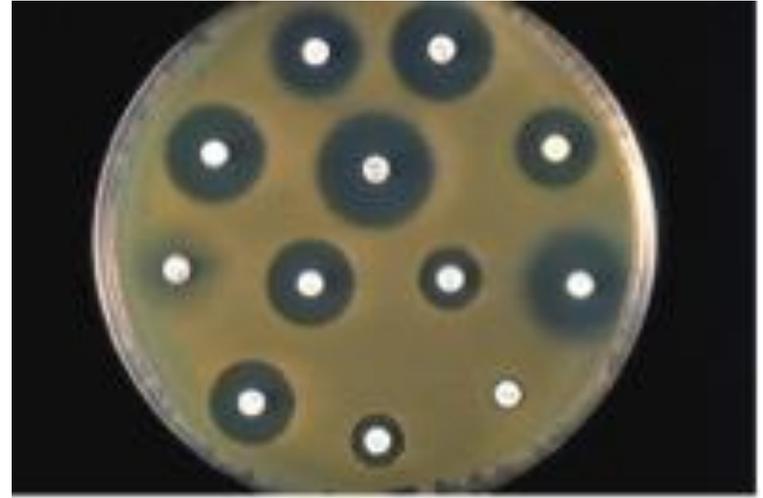
Accelerate Diagnostics - Pheno System

- Antimikrobiale duyarlılıklarının belirlenmesi

4-5 saat



24-48 saat



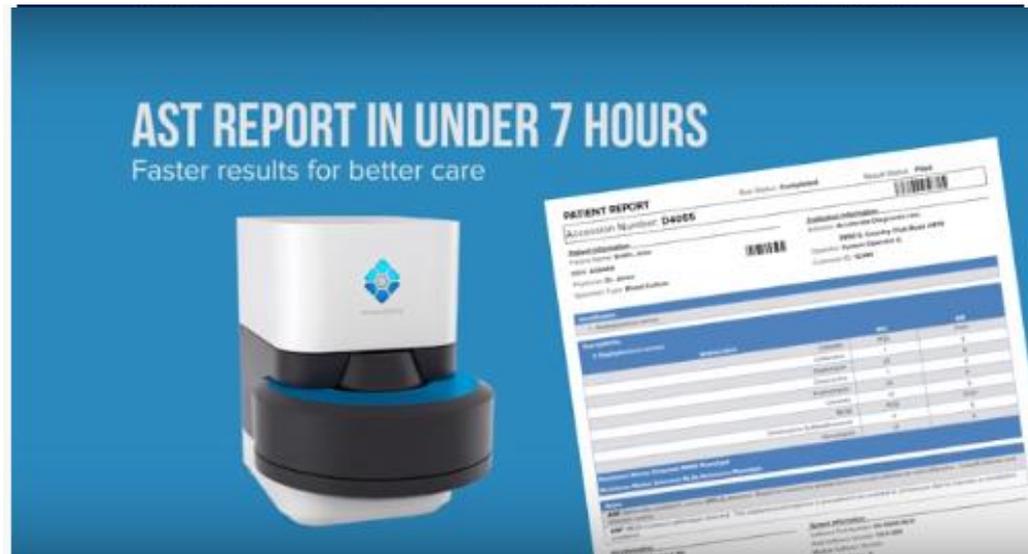
Accelerate Diagnostics - Pheno System

- Antimikrobiale duyarlılık testi; zaman-atlamalı mikroskop ile **üreme/inhibisyonun** takibi



Accelerate Diagnostics - Pheno System

- Antimikrobiale duyarlılık testi; zaman-atlamalı mikroskop ile **üreme/inhibisyonun** takibi



Accelerate Diagnostics - Pheno System

ORIGINAL ARTICLE

Rapid Automated Microscopy for Microbiological Surveillance of Ventilator-associated Pneumonia

Ivor S. Douglas¹, Connie S. Price², Katherine H. Overdier¹, Robert F. Wolken³, Steven W. Metzger⁴, Kenneth R. Hance⁴, and David C. Howson⁴

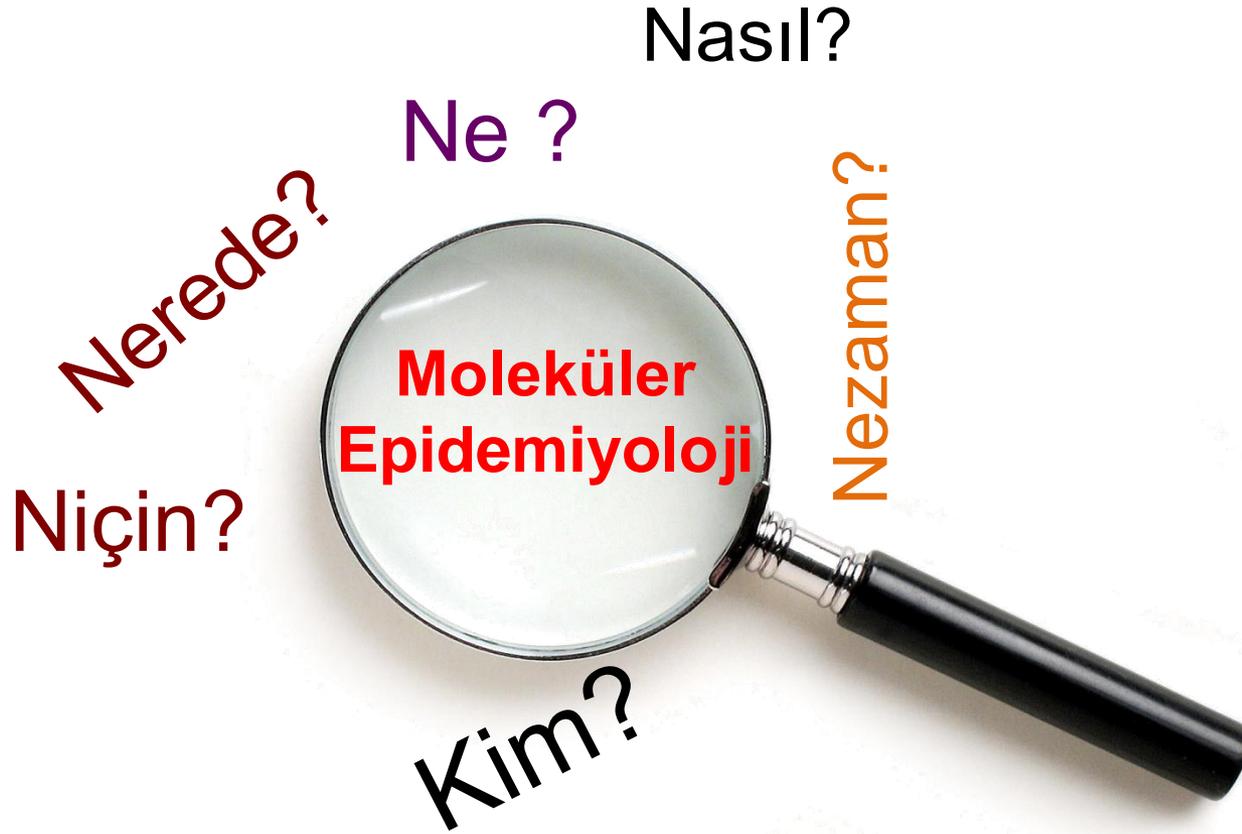
¹Division of Pulmonary Sciences and Critical Care Medicine, ²Division of Infectious Disease, and ³Respiratory Therapy, Department of Medicine, Denver Health Medical Center, Denver, Colorado; and ⁴Accelerate Diagnostics, Inc., Tucson, Arizona

		CLINICAL MICROBIOLOGY PRESENCE/ABSENCE $\geq 1 \times 10^4$ CFU/mL	
		Positive	Negative
AUTOMATED MICROSCOPY	Positive	7	2
	Negative	0	64

Sensitivity=100% Specificity=97%

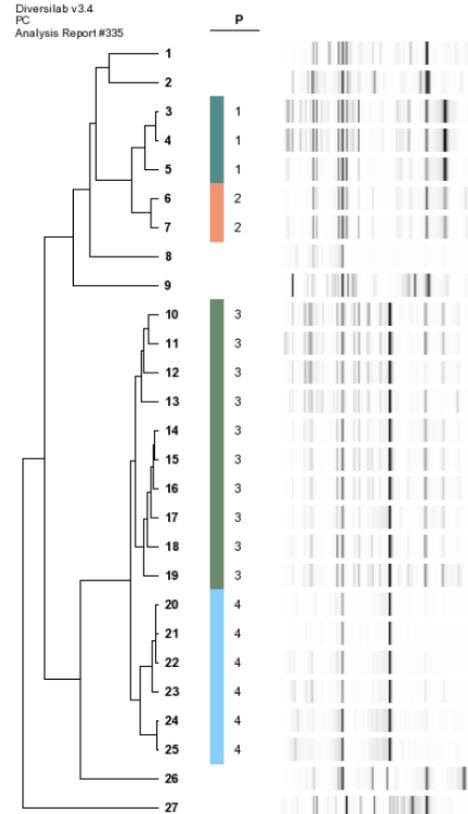
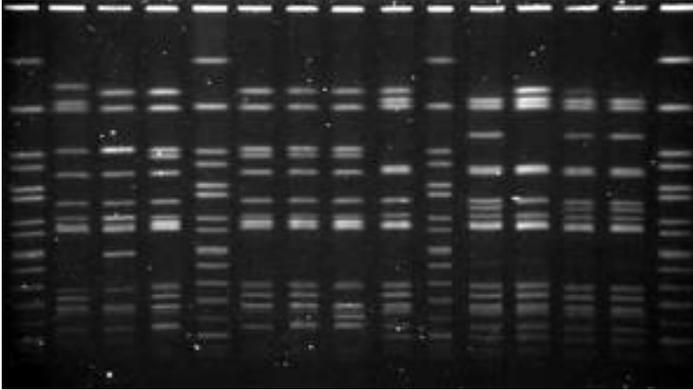
Figure 1. Performance characteristics of automated microscopy compared with clinical culture for qualitative presence or absence of ventilator-associated pneumonia (VAP)-associated bacteria above the VAP diagnostic threshold of 1×10^4 colony-forming units (cfu)/ml.

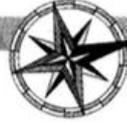
Moleküler Epidemiyoloji



Moleküler Epidemiyoloji

- Bugün için ise moleküler epidemiyoloji terimi; genellikle “moleküler biyolojik yöntemlerle suş tiplendirme” çalışmalarına verilen ortak bir isim gibi kullanılmaktadır.





Population Biology, Evolution, and Infectious Disease: Convergence and Synthesis

Bruce R. Levin,¹ Marc Lipsitch,¹ Sebastian Bonhoeffer²

Traditionally, the interest of population and evolutionary biologists in infectious diseases has been almost exclusively in their role as agents of natural selection in higher organisms. Recently, this interest has expanded to include the genetic structure and evolution of microparasite populations, the immune response, and the population genetics of medical and public health interventions. In these areas, emphasizing the ways in which these approaches have been contributing to the design and evaluation of interventions.

Moleküler epidemiyolojinin sınırlarını;

- etken mikroorganizmanın tespiti
- bulaşma yollarının araştırılması
- virülanslarından sorumlu genlerin belirlenmesi ve
- izolatlar arası klonal ilişkilerin araştırılmasını kapsayan geniş bir yelpazede çizilmiştir.

Moleküler Epidemiyoloji

- Hastane infeksiyonuna neden olan **etkenlerin izini sürebilmek için;**
 1. **izolatların moleküler karakterizasyonu yapılmalı ve**
 2. **izolatlar arasındaki klonal ilişkiler genotiplendirme yöntemleriyle araştırılmalıdır.**

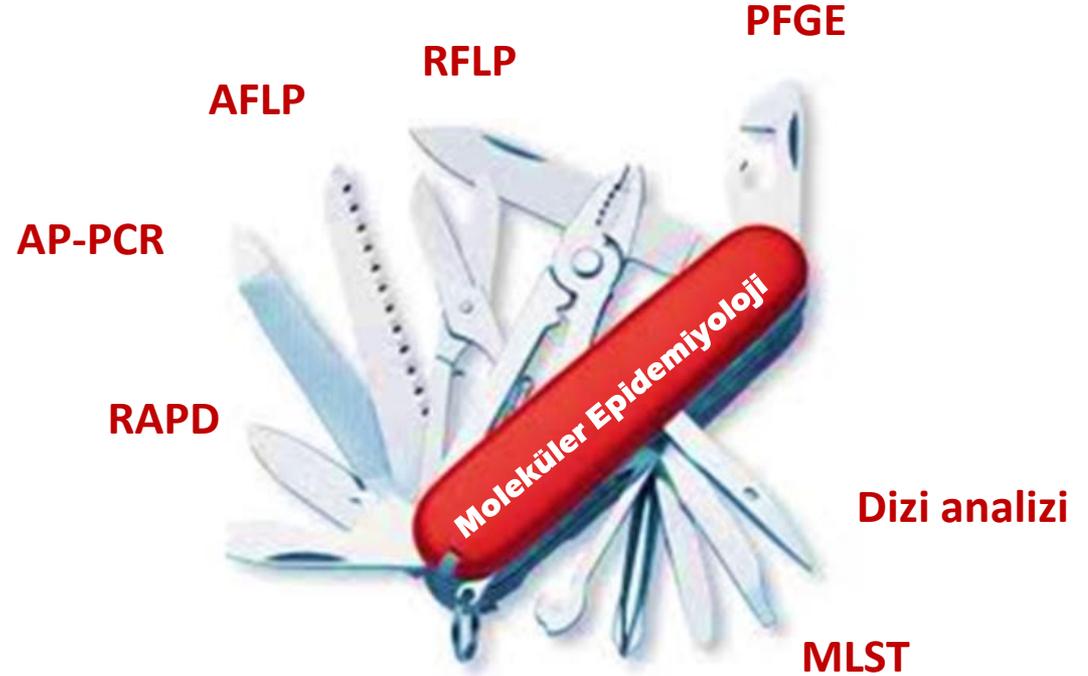
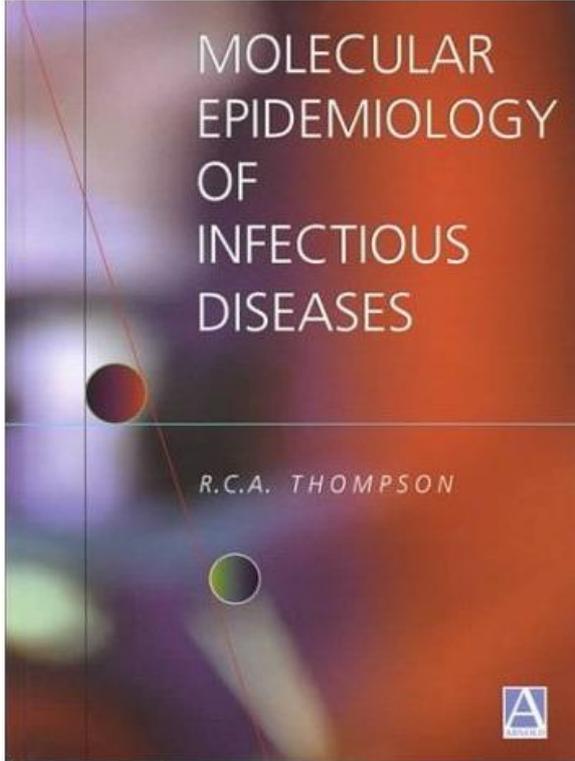


İzolatlarda Moleküler Karakterizasyonu



- İzole edilen mikroorganizmaların moleküler karakterizasyonu için;
 - antimikrobiyal direnç genleri
 - virülans genleri
 - mobil genetik elemanları
 - plazmid
 - transpozon
 - İntegron
 - hücre duvar yapıları araştırılmalıdır.

İzolatlar Arasındaki Klonal İlişkilerin Araştırılması



Moleküler epidemiyoloji için yöntemler

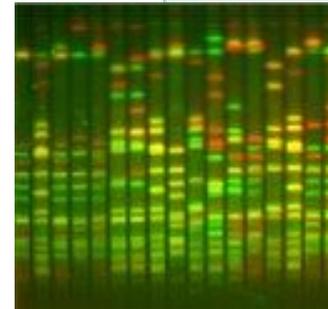
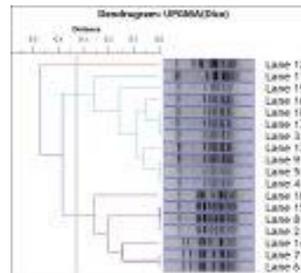
- Hastane infeksiyonları ile ilişkili mikroorganizmaların moleküler tiplendirilmesinde; **kromozomal DNA polimorfizmine dayalı olan yöntemler**, hızlı ve güvenilir olmalarıyla ön plana çıkmaktadırlar.

- *Pulsed field gel electrophoresis (PFGE)*
- *PCR-restriction fragment length polymorphism (PCR-RFLP)*
- *Random amplified polymorphic DNA (RAPD)*
- *Arbitrarily primed polymerase chain reaction (AP-PCR)*
- *Repetitive extragenic palindromic element-PCR (rep-PCR)*
- *Amplified ribozomal DNA restriction analysis (ARDRA)*



Pulsed Field Gel Electrophoresis (PFGE)

- Sonuçların analizi ve yorumlanması
 - Bu amaçla en çok kullanılan bilgisayar programları;
 - Gel-Compar (Applied Maths, Kortrijk, Belgium)
 - Dendron (Solltech, Oakdale, Iowa)
 - Diversity Database Fingerprinting Software (Bio-Rad)
 - Gene Profiler (Scanlytics, Fairfax, Va)
 - Phoretix 1-D (Nonlinear USA, Durham, N.C.)
 - Quantar Pro (KeyGene Products, Wageningen, Netherlands)
 - Taxotron (Taxolab, Institut Pasteur, Paris, France)



Pulsed Field Gel Electrophoresis (PFGE)

- Sonuçların analizi ve yorumlanması



Moleküler Epidemiyoloji Ne Zaman Kullanılmalı ?

• Her zaman

NEW MICROBIOLOGICA, 31, 401-408, 2008

Infection

Correspondence

Epidemiological Characteristics of Fatal *Candida krusei* Fungemia in Immunocompromised Febrile Neutropenic Children

In early 1990s, fluconazole was introduced as a prophylactic antifungal agent after stem cell transplantation (SCT). At the same time, *Candida krusei* emerged as a chief fungal pathogen among the patients with SCT [1, 2]. In our SCT center, which has a 15-bed capacity unit dedicated for children with malignant diseases and 5-bed unit for SCT patients, low-dose fluconazole is routinely included in the posttransplant prophylaxis regimen (starting at day 7). In recent years, we observed mortality due to fungal sepsis in patients undergoing allogeneic SCT and chemotherapy. Here, we report the characteristics of the patients having mortal *C. krusei* fungemia and the molecular typing results of the *C. krusei* strains in order to develop more effective policies for treatment and prevention of fungal infections.

The febrile neutropenic episodes of the 47 children (median age: 9.13 ± 4.73 years) were prospectively evaluated from March 2001 to January 2003. For each patient, daily leukocyte counts and body temperatures were measured. Fever was defined as a single oral temperature of ≥ 38.5°C or a temperature of ≥ 38.0°C for ≥ 1 h. Neutropenia was defined as a neutrophil count of < 500 cells/mm³, or a count of < 1,000 cells/mm³ with a predicted decrease to < 500 cells/mm³ (Guidelines from the Infectious Disease Society of America, 1997). Patients who had both fever and neutropenia were recognized as febrile neutropenic. Nine patients developed severe fungal infection during neutropenia. Three of those having bloodstream infection with the *C. krusei* were analyzed in detail, in this study.

Surveillance cultures were taken from blood, throat, stool, urine and catheter exit sites of the patients in the SCT unit when they had febrile episode and/or weekly on a routine basis. Automated blood culture system (Becton Dickinson, pediatric stals, USA) and BACTEC MYCO/F LYTC (Becton Dickinson) were used for blood cultures. To search exogenous sources of *C. krusei* sepsis, 51 specimens from the hands of 32 health care workers (9 doctors, 11 nurses, 4 laboratory workers, and 8 cleaning staff) and 19 environmental swabs from commonly used areas around the patients (nurse desk, room and toilet doors, refrigerator doors, telephones, etc) were analyzed.

antifungal susceptibility. Molecular typing of the nine *C. krusei* strains of the three patients (three blood, two stool, one urine, one throat, one vagina, and one sputum isolates) was performed by pulsed field gel electrophoresis (PFGE) following the protocol described previously [3].

A total of 156 febrile episodes were recorded in the 47 patients within the 312 febrile neutropenic days. Bacteria were isolated in 28 (52%) and fungi in 26 (48%) of the 54 clinical specimens. It was noted that the patients with prolonged and deep neutropenia (hematological malignancy patients) are under an increased risk for fungal infections [1, 4, 5]. In agreement with these findings, we observed systemic fungal infections with high mortality in recent years. Of the 47 patients with febrile neutropenia, 14 had fungal isolation from various body sites and 9 (19%) died due to fungal infection. Eight of these fatal cases had non-*Candida albicans* infection (Table 1). Distribution of *Candida* spp. among the five non-fatal cases was as follows: *C. krusei* (1), *C. kefyr* (1), *C. albicans* (2), *C. lusitana* (1). In an Italian tertiary hospital, it was found that 60% of the candidemia episodes were related to non-*albicans* *Candida* and high mortality rates were observed particularly for hematological (71%) and transplant patients (50%) [6]. In another tertiary care hospital in

Infection 2008; 36: 88-91
DOI 10.1093/infdis/jin00742461

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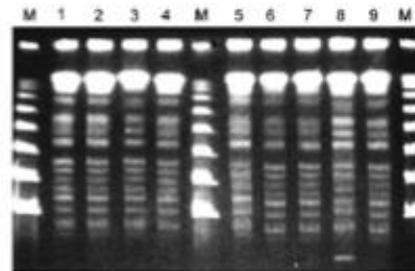
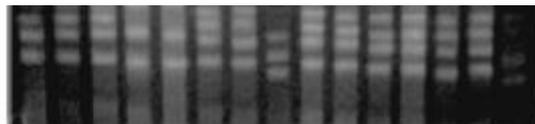


Figure 1. PFGE profile of the 9 *C. krusei* strains. M: molecular weight marker. Lanes 1 and 2: patient 1's blood and stool isolates; lanes 3-6: patient 2's throat, stool, blood, and sputum isolates, respectively; lanes 7-9: patient 3's blood, vagina, and throat isolates, respectively.

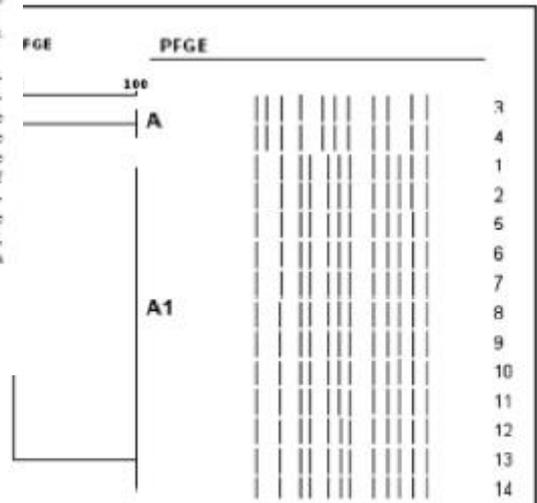
breaks lasting 1 or 2 days or ineffective doses due to difficulties to obtain liposomal amphotericin B. Amphotericin B treatment was started at tenth day of the febrile neutropenia for two patients and at the first day of the febrile neutropenia and sepsis for the third patient. These patients died on days 8, 21, and 23 after the diagnosis of sepsis. Since there were no concomitant bacterial infections in these patients having many risk factors, the mortality was attributed due to *C. krusei* sepsis. However, it is possible that the high mortality in these patients was

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CA, 30, 131-137, 2007

C. aeruginosa
infection
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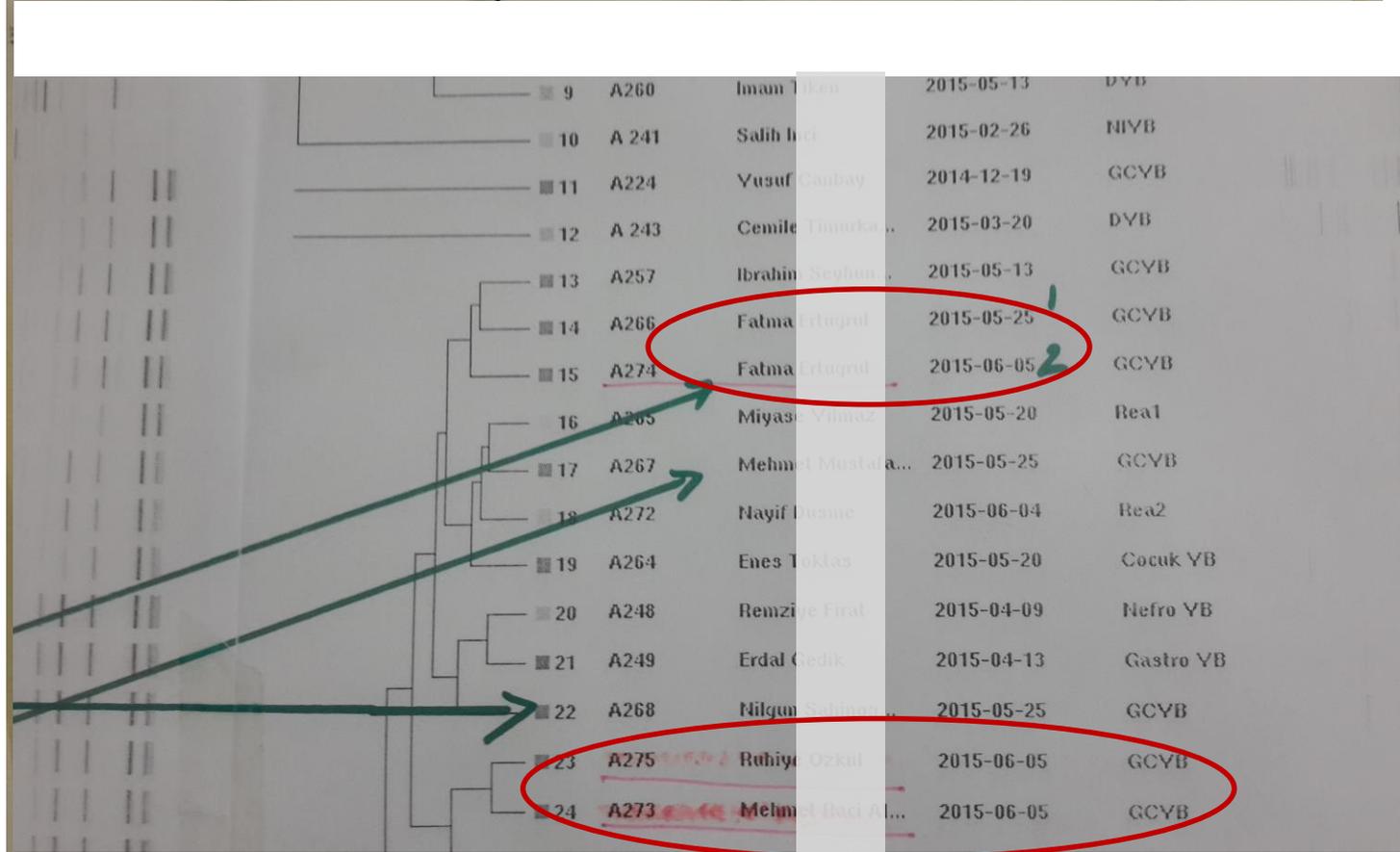
AD, Yasemin Enay, MD, *



Moleküler Epidemiyoloji Ne Zaman Kullanılmalı ?



Moleküler Epidemiyoloji Ne Zaman Kullanılmalı ?



Eldeki tüm imkanlar kullanılmalı

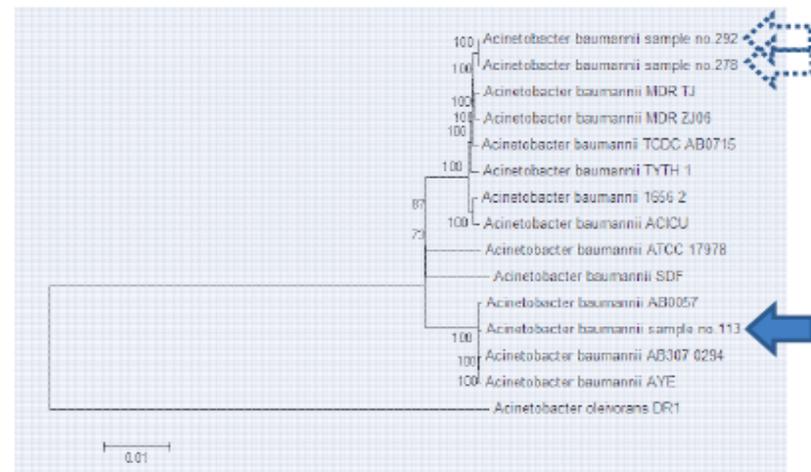
Diagnostic Report: *Acinetobacter baumannii*

Isolate number	113	Date isolated	2012-09-04
Origin	Throat swab	Pat. ID	XXXX-XXGiXXX

MLST type

1

Phylogeny



Resistance genes	Antibiotic group	Location
<i>bla</i> _{oxa-72}	Beta-Lactam	Plasmid 1
<i>bla</i> _{oxa-66}	Beta-Lactam	Chromosome
<i>aadA1</i>	Aminoglycoside	Chromosome
<i>sul1</i>	Sulfonamid	Chromosome

Virulence genes detected	Location
Type IV secretion system	Plasmid 2

Assessment

CAVE: Strain with two carbapenemase genes detected. Classified as **multiresistant**. Phylogeny relation traced to known outbreak strain AB0057. No significant relation to other clinical isolates in recent time frame.

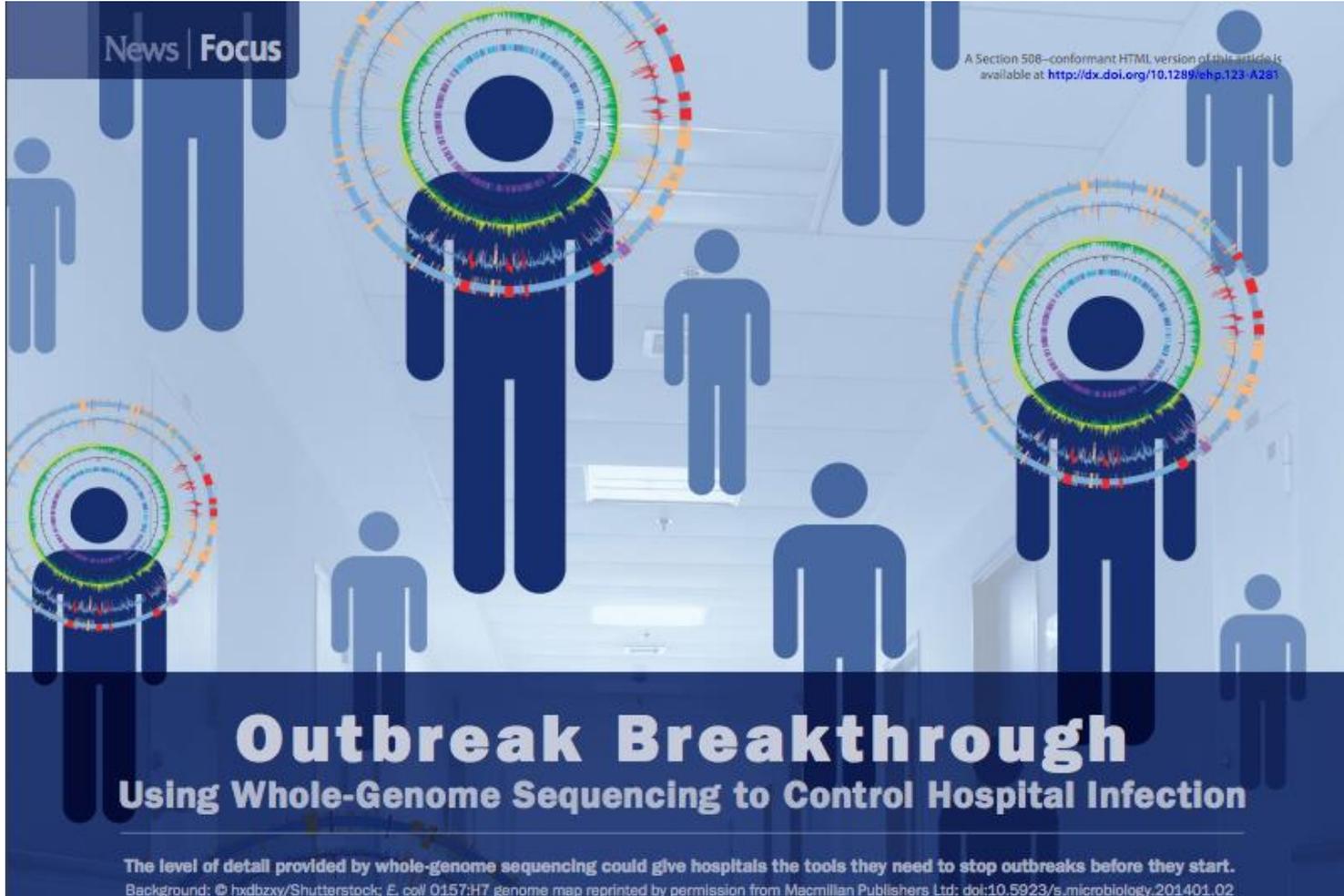
Hygiene: Single patient isolation recommended.

Treatment options: Tetracyclin, Tigecyclin, Colistin (inhalative)



Eldeki tüm imkanlar kullanılmalı

- *Yeni Nesil Dizileme Sistemleri*



News | Focus

A Section 508-conformant HTML version of this article is available at http://dx.doi.org/10.1289/ehp.123_A281

Outbreak Breakthrough

Using Whole-Genome Sequencing to Control Hospital Infection

The level of detail provided by whole-genome sequencing could give hospitals the tools they need to stop outbreaks before they start.

Background: © hxdzxy/Shutterstock; *E. coli* O157:H7 genome map reprinted by permission from Macmillan Publishers Ltd: doi:10.5923/s.microbiology.201401.02

Eldeki tüm imkanlar kullanılmalı

Broad Active Surveillance Enabled by Pathogenica's HAI BioDetection Kit

Jack T. Leonard, Adeyemi Adesokan, Stephen Brusco, Thomas Clarke IV, Graeme Doran, Sarah Gruszka, Sarah Mahoney, and Alexander Rolfe

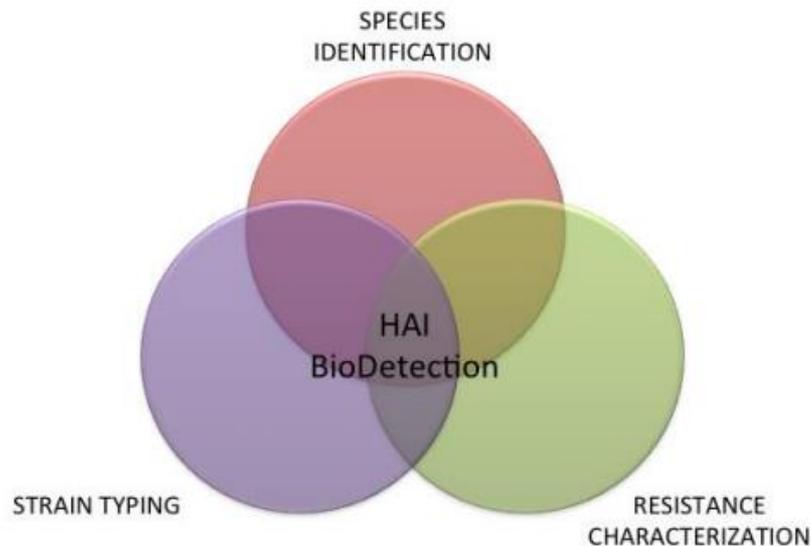
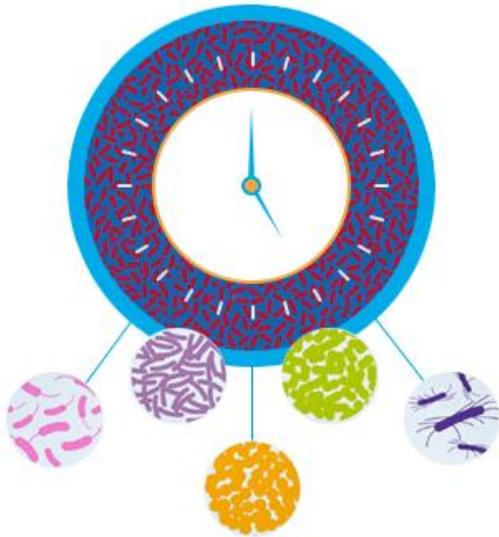


Figure 1: In addition to active surveillance, HAI BioDetection is suitable for strain-typing and antimicrobial resistance characterization.

Eldeki tüm imkanlar kullanılmalı

bioMérieux EpiSeq™* :
When Next-Generation Sequencing meets
microbiology serving epidemiology

* Not for diagnostic use



bioMérieux EpiSeq™, the service of choice to help manage Healthcare-Associated Infections (HAI) using Next-Generation Sequencing (NGS) technology

Fighting Healthcare-Associated Infections is your critical challenge so put the power of Next-Generation Sequencing on your side.

Moleküler Epidemiyoloji Ne Zaman Kullanılmalı ?

- Hastane infeksiyon kontrol komitesine **anlık bilgi verdik** ve haftalık toplantılarına katıldık.
- Yoğun bakımlarda salgınlardan **etkilenen hasta sayısı artmadan** müdahale edilmesine katkıda bulunduk.
- Hastane infeksiyon kontrol komitesinin **elini güçlendirdik**.

